

Community composition predicts photogrammetry-based structural complexity on coral reefs

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complexity on coral reefs 2 3 J. Carlot^{1-2*}, A. Rovère³⁻⁴, E. Casella³⁻⁴, D. Harris⁵, C. Grellet-Muñoz¹⁻³, Y. Chancerelle¹⁻², 4 5 E. Dormy ⁶, L. Hedouin¹⁻², V. Parravicini¹⁻² 6 7 ¹PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRIOBE, BP 1013, 98729 Papetoai, 8 Moorea, French Polynesia 9 ²Laboratoire d'Excellence « CORAIL » 10 ³Centre for Marine Environmental Sciences (MARUM), Bremen University, Bremen, 11 Germany. 12 ⁴Leibniz Centre for Tropical Marine Research, Bremen, Germany. 13 ⁵The University of Queensland, School of Earth and Environmental Sciences, Brisbane, 14 Oueensland, Australia. 15 ⁶Department of Mathematics and Applications, CNRS UMR 8553, Ecole Normale Supérieure, 16 Paris, France 17 18 **Keywords** 19 Coral Complexity – Rugosity measures – Photogrammetry – Reef ecology – Modeling 20 21 *Corresponding author: Jérémy Carlot, 22 PSL Research University 23 **EPHE-UPVD-CNRS** 24 USR 3278 CRIOBE 25 BP 1013, 98729 Papetoai, Moorea, French Polynesia 26 Laboratoire d'Excellence « CORAIL »

Community composition predicts photogrammetry-based structural

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Abstract

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The capacity of coral reefs to provide ecosystem services, to keep their diversity and their productivity are related to their three-dimensional structural complexity. This parameter is also correlated to total fish biomass, to the general reef resilience to external stresses and to their ability to dissipate wave energy. However, information on structural complexity (also defined as reef rugosity) has been uncommonly assessed in historical monitoring programs, with the result that the long-term trend of this variable is generally unavailable. In this study, we show that it is possible to predict and hindcast the three-dimensional complexity of coral reefs by combining photogrammetry, statistical modeling and historical benthic community data. We calibrated a lasso generalized linear model to predict structural complexity obtained by 57 photogrammetry transects recorded at 13 sites around the island of Moorea (French Polynesia). Our model was able to predict structural complexity with high accuracy (crossvalidated $R^2 = 0.81 \pm 0.12$). We then used our model to hindcast historical trends in 3D structural complexity using community composition data collected in Moorea from 2004 to 2017. The temporal analysis highlighted the dramatic effect of a crown-of-thorns outbreak in 2006-2009 and Cyclone Oli in 2010. These two events together reduce coral cover from ≈50% to almost zero. Our model captured these effects, confirming the possibility to predict structural complexity on the basis of assemblage composition.

Introduction

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Global concerns are emerging about the increasing frequency of mass mortality of corals associated to coral bleaching events (Van Oppen and Lough 2009; Heron et al. 2016; Hughes et al. 2017). These disturbances are associated to severe habitat destruction that reduces the structural complexity (i.e., flattening) of coral reefs (Newman et al. 2015). Structural complexity is the three-dimensional spatial arrangement of an ecosystem (McCormick 1994; Chazdon 2014), which is largely due to the growth form and distribution of hard coral. According to the habitat heterogeneity hypothesis (MacArthur and Wilson 1967), the more complex the structure of an ecosystem, the greater the diversity and abundance of associated organisms. On coral reefs, the 3D structural complexity of the habitat is correlated to the biomass and diversity of fish (Willis and Anderson 2003; Gratwicke and Speight 2005; Alvarez-Filip et al. 2009; Rogers et al. 2014), to the reef capacity to recover from disturbance (Graham et al. 2015), but also to the reef ability to dissipate wave energy, thus protecting the shoreline from extreme inundations (Harris et al. 2018). Broad-scale declines in the complexity of coral reefs have been observed both in the Caribbean and the Pacific as a result of both human impacts and climate changes (Hoegh-Guldberg 1999; Hughes et al. 2003; Hoegh-Guldberg et al. 2007; Perry et al. 2018). Despite the well-known important relationship between structural complexity, ecological diversity, abundance and biomass, information on structural complexity is sparse in monitoring programs with the result that long-term trends for this variable are virtually unknown (Graham et al. 2015). Methods to measure structural complexity (often referred as rugosity, in particular in older literature) on coral reefs first arose in the 70' in articles by Risk (1972) and Hobson (1972). These authors defined three criteria for measuring complexity: 1) the measure had to be easily understandable, 2) it had to be measurable during the fieldwork and, 3) it should be comparable. In early studies, it was proposed that rugosity could be recorded by draping a steel

chain over the reef surface, and then measuring the ratio between the total length of the chain

and the planar distance between the ends of the chain. The higher the ratio, the more complex the substratum (Hill and Wilkinson 2004; Graham and Nash 2013). Despite the ease of use of such metric, laying a chain represents a bi-dimensional measure which does not capture the full complexity of complex three-dimensional (3D) habitats such as coral reefs. Although some time-consuming 3D metrics have been proposed in the past (e.g. Parravicini et al. 2006), the recent progress in underwater photogrammetry are finally affording researchers the opportunity to capture the three-dimensionality of coral reefs. For example, Friedman et al. (2012) started to use a georeferenced survey work and each includes a downward-looking camera pair with a baseline of approximately 7cm, pixel resolution of 1360 × 1024 to define rugosity. Others authors like Burns et al. (2015) were using these new advances for defining the rugosity about a transect and extract a complexity index at the species level for 6 species. Leon et al. (2015), defined three roughness parameters, namely the root means square height, tortuosity (i.e. rugosity) and fractal dimension, and were derived and compared in order to asses which one better characterizes reef flat roughness. Naughton et al. (2015) succeed with an accuracy never equaled to define maps of community structures between taxa. Some writers have even pushed the boundaries to measure the small-scale three-dimensional features of a shallow-water coral reef thanks to drone (Casella et al. 2017). Thus, a plethora of works have emerged asking several authors about the chain-tape future (Storlazzi et al. 2016). However, whatever the metric employed (chain-tape or photogrammetry methods), there is still no clear evidence concerning what is driving structural complexity. Some authors claim that it is driven by the presence of some branching species like the Acropora spp. and thus the overall coral cover would not matter (Aronson and Precht 2006; Alvarez-Filip et al. 2009, 2011). Others have found that coral cover is significantly and highly correlated to the rugosity (Halford et al. 2004; Graham and Nash 2013) or species composition (Richardson et al. 2017). But in both cases, there is a common consensus for admitting coral drives complexity which could be used for rebuilding past rugosities.

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In this study, we combined statistical modeling to 3D reef transects reconstructed using photogrammetry in order to test the potential to predict coral reef structural complexity on the basis of benthic community composition. We study the reefs of Moorea (French Polynesia) where, using benthic communities time series we back-calculate reef structural complexity. We were able to retrace two relevant episodes of habitat destruction: the Acanthaster planci outbreak of 2006-2009 and cyclone Oli of 2010.

Material and Methods

1. Study area

Moorea is a tropical volcanic island of volcanic origin, located in the Pacific Ocean between 17.4714° and 17.6058° South and 149.7522° and 149.9269° West. The island is shaped as a triangle with a perimeter of 61 km and coastlines facing north, southwest and southeast (**Fig. 1**). The island is encircled by a coral reef, that is 500 to 700 m wide; the only exception to this pattern is the Northeast extremity where the lagoon width is limited to few tens of meters. Moorea is exposed to Northwest winds from January to March. Tides are semidiurnal with an amplitude of less than 0.3 m (Chazottes et al. 1995; Leichter et al. 2013). The swell direction is from southwest to northeast during the whole year.

The reefs of Moorea are threatened by several disturbances of biotic and abiotic origin (Adjeroud et al. 2018). The most devastating biotic disturbances were the 1979 and 2006 Acanthatster planci outbreak, which reduced coral cover from 50% to 10% (in 1979; Berumen and Pratchett 2006) and from 50% to less than 10% (in 2006; Lamy et al. 2016). Abiotic disturbances also impacted Moorea island, with main cyclones recorded in 1991 (Wasa) and 2010 (Oli). The impact of Wasa reduced the coral cover by 5% to over 20% across all the island. In contrast, coral cover was reduced to lower than 5% by Cyclone Oli (Lamy et al. 2016; Adjeroud et al. 2018) (**Fig. 2**).

2. Rugosity measures

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In Moorea, a total of 57 photogrammetric transects were surveyed in end-of-2015 – beginning-of-2016, at three different sites: Tiahura (North Coast, 21 transects); Haapiti (Southwest coast, 20 transects) and Temae (Southeast coast, 16 transects) (Fig. 1). Each transect consisted of swaths of 20m length and 2m width and all transects were carried out in the outer reef, between 5 and 8m depth. Each transect was set up fixing on the bottom a 2m-long chain and, perpendicularly to it, a 20m-long metered tape. A diver swam ≈2m above the sea-bottom, maintaining the swimming speed as constant as possible and collecting images with a GoPro Hero camera pointed towards the sea-bottom. The camera was set to collect photos (12 megapixels) in time-lapse mode (2 pictures per second). For each transect, we collected approximately 200 photos with a forward overlap of ≈90%, with the diver swimming over the length of the entire transect four times to allow optimal side overlap. After the collection of the photos, the diver noted the depth of each extremity of the chain and metered tape to use them in the photogrammetric process as Ground Control Points (GCPs). In the case of an on-the-job self-calibration, the camera calibration is derived from image coordinates measured in the mapping photography and including the camera calibration parameters as unknowns in a selfcalibrating bundle adjustment (Harwin et al. 2015).

The set of photos and the GCPs collected were then used as input to Agisoft Photoscan (www.agisoft.com), a photogrammetry software based on the Structure from Motion (SfM) method (Ullman 1979; Westoby et al. 2012). We used Agisoft to build the orthophotomosaic and the Digital Elevation Model (DEM) of each transect, with the same procedure explained by Storlazzi et al. (2016). Details of the photogrammetric process are shown in the **Annex S1**. Subsequently, we imported the DEM in ArcGIS v10.2 and calculated the reef rugosity by dividing the surface of the DEM area by the area of its planar projection (approximately 40m^2) (**Fig. 3**). For the 57 transects, we estimated an average horizontal error of $0.1\pm0.06\text{m}$ and average vertical error of $0.04\pm0.04\text{m}$, internal to the reconstructed model. Only the vertical error

had an influence on the estimation of rugosity, but given the range of the error, it was considered negligible.

3. Benthic community description and assessment

The orthophotomosaics were imported into Coral Point Count, which was used with an Excel extension v4.1 (Kohler and Gill 2006). We assessed the benthic cover placing 100 random points on the photomosaic and described 8 distinct benthic cover categories (**Table 1**). A second dataset was used according to the CRIOBE surveys from 2004 to nowadays and consists of 25m point intercept transects data collected at 13 sites around the island across three habitats (fringing reef, back reef, outer reef) and using the same benthic categories (**Fig. 1**). This classification is based on the guideline of the monitoring program created by SO CORAIL (http://observatoire.criobe.pf/CRIOBEData/), the coral reef observation program of the French National Institute for Earth Sciences and Astronomy (INSU). These categories enabled us to back-calculate structural complexity on time series data from Moorea. For matching both datasets according to the habitat, only the outer reef was selected.

4. Statistical analysis

Our main goal was to calibrate a model that predicts structural complexity according to the benthic community composition. Since benthic community cover is expressed as percentages, we preferred not to use them as predictor variables as they are heavily correlated, and collinearity would have been high. We preferred to build a database that include the time series data and our photogrammetry transects without transforming the data to perform multivariate analysis. Thus, the Euclidean distance could be used to conduce a Principal Component Analyses (PCA). The first five orthogonal axes (accounted for more than 75% of the variance) were then extracted to be used as predictor variables in the model. A lasso generalized linear model has been conducted to predict coral reef structural complexity

according to the 5 PCA axes. A 10-Fold cross validation was done and a step AIC procedure defining the best model was applied. Because the 57 transects were measured at the same depth, those transects were pooled together rather to apply a mixed model due to the lack of the data, making us benefit from a solid robustness. The best model selected kept only 3 PCA dimensions. According to the k-fold analysis, k R² were obtained giving a necessary uncertainty for the model. This method appears more meaningful estimate than classical R² when the model has to be used for predictions. The lasso generalized linear model was then applied to time series data to back-calculate structural complexity and to test whether our model was able to detect drops of structural complexity due to the major perturbations.

The PCA data for the time series in Moorea were used to produce PCA plots and define the long-term tendencies in order to descriptively assess the entity of the effect of the Acanthaster planci outbreak (2006-2009) and the Oli cyclone (2010) on benthic communities. An ANOVA was then applied with the aim to define several bunches of similar years according to the PCA axes. To match those years with each other, a Tukey post-hoc analysis was conducted and a matrix of results was elaborated. Finally, special attention has been given about the Acropora spp. cover, Pocillopora spp. cover and rugosity index in 2004 (pre-disturbance year, with the high percent cover of corals) and 2017 (post-disturbance year, with the high percent cover of corals) to define a potential resilience or recovery (**Table S1**).

Results

The analysis of time series revealed that coral diversity was higher in 2004 with a percent cover of corals (CC) of 44.08% (**Fig. 4**). The CC decreased from 2004 to 2010 down to a minimum of 3.62%, which corresponds to the event of Cyclone Oli. After the cyclone passed, the coral cover increased over time until the end of the series (2017) with a final value of 42.77%. In 2004 the coral reefs of Moorea also showed a greater diversity of coral morphology (massive, branching in general, columns and encrusting). The assemblages

remained fairly stable despite a slow decline of the CC the first 2 years ($CC_{2004} = 44.08\%$ to $CC_{2006} = 40.62\%$). Then, a first Crown-Of-Thorns Starfish (COTS) outbreak was reported in early 2006 (Kayal et al. 2012) and continued until 2009. The following year, cyclone Oli hit the island further decreasing the CC. After these events and until 2014, the substrate consisted of rubble and cobbles. From 2015, the CC recovered to a state similar to that of 2004. However, compared to 2004, the coral cover in 2017 was dominated by Pocillopora spp. (20.10 \pm 6.78% in 2004 vs 26.61 \pm 14.52% in 2017) instead of a more diverse assemblage with a high abundance of Acropora spp (9.76 \pm 5.61% vs 2.53 \pm 1.90% in 2017).

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The cross-validated R² (CV-R²) from our model reaches 0.81 ± 0.12 . The 3 dimensions used were all significant and non-correlate (Table 2, Fig S1). The back calculation of structural complexity captured these major shifts in community structure. High structural complexity was observed in 2005 and 2006. All sites were then predicted to lose complexity corresponding to the timing of COTS outbreak models present patterns of decrease and increase in rugosity, matching with the biotic and abiotic changes like the COTS outbreak and the Cyclone Oli, as discussed above (Fig. 5). The model also exemplifies that the resilience of structural complexity differs among the thirteen reefs studied around Moorea island. In 2004, the highest values of structural complexity were measured respectively at Pihaena (North), Motu Ahi (East) and Haapiti (Northwest) (3.86, 3.51 and 2.92 respectively) while the lowest were recorded in Maatea (Southeast), Tiahura (North) and Aroa (North) (2.26, 2.07 and 1.81 respectively). After the disturbances in 2010, the higher values were defined in Taotaha (Northwest), Afareaitu (Southeast) and Entre 2 Baies (North) (2.08, 2.10 and 2.10 respectively) while the lowest values were documented at Haapiti (Northwest), Tiahura (North) and Maatea (Southeast) (1.84, 1.66 and 1.15 respectively). Finally, in 2017, the 3 lowest values were measured on the East coast (Motu Ahi, Maatea and Temae with rugosity values of 2.35, 2.12 and 1.98 respectively) while the sites located on the northwest side (Entre 2 Baies, Tiahura and Gendron) presented the 3 higher rugosity scores (4.36, 4.31 and 3.53 respectively).

Finally, rugosity values rise to values equivalent to the first year of monitoring, during the last year of monitoring in 2017. The presence of Acropora spp. shows a significant difference between 2004 and 2017. However, no significant difference could be observed in either the Pocillopora spp. presence or the rugosity score (**Fig. 6**). In addition, the ANOVA followed by the post-hoc analysis confirmed a difference between two bunches of years according to the complexity around the island. Indeed, two profiles were highlighted: 1) one from 2004 to 2007 and 2016 to 2017 and 2) one from 2008 to 2015. These results support that the 3D complexity came back to an equilibrium four years after the Oli cyclone (**Table S1**).

Discussion

In this study we used a combination of methods – coral reef photogrammetry and statistical models – to test the potential to use species composition data to predict the structural complexity of coral reef assemblages. The use of the photogrammetry allowed us to obtain a three-dimensional metric of structural complexity, compared to linear metrics classically used such as the chain transect (Burns et al. 2015). Photogrammetry permits us to cover 40m^2 of the reef in one dive of about 90 minutes, whereas the chain method usually requires approximatively 15 minutes for a simple 20m transect. However, even if the pixel size of 1.73 \times 1.73 μ m - used for defining the complexity - is higher resolution than what is achieved using chain and tape, the results have to be interpreted with cautious. Indeed, a number of studies have shown photogrammetry to be error prone in a number of different ways. Lavy et al. (2015) and Figueira et al. (2015) both found that branching corals and other complex growth forms produce more error in photogrammetry-based estimates of complexity compared to in situ methods. Furthermore, Bryson et al. (2017) found that environmental conditions,

postprocessing, and even taking photos underwater can impact the accuracy of 3D structure estimates using photogrammetry. In addition, this model is relevant for planar parts of the reef, however, facing dropoff would be challenging. This technique requires swimming over the bottom, nevertheless, as the more the distance you add between you and the bottom, the worst the reconstruction will be. Finally, all the hidden part (not present in the photos) are not reconstructed (e.g. what is inside a hole). The latter limit causes relatively heavy consequences to define an accurate measure of the rugosity in those conditions. From a statistical point of view, Carroll et al. (2006) defined 3 different regimes of swell in Moorea which could altered the benthic composition, according to the exposure due to the side of the island. Even if these affirmations are directly observable concerning the rugosity in 2017 with our current model (lowest values on the east coast and higher values on the north coast) a mixed model would have been more relevant. Unfortunately, only 16 transects were done on the southeast side of Moorea which is not allowed us for using a mixed model. The robustness of the model would have been directly impacted by the lack of residuals (rugosity values sometimes lower than 1; Launer & Wilkinson, 2014). As per any statistical model, the accuracy of prediction will increase with the size of the calibrating dataset. In that context, more data will likely be needed to accurately capture spatial variation.

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Despite these limits, we have found a significant relationship between reef structural complexity and the composition of the benthic assemblages. Indeed 3 PCA axes were enough to accurately predict complexity with a high accuracy (CV-R² = 0.81 ± 0.12). To validate our model, we compared our values to (Kayal et al. 2017). These authors have found values at 12m-depth of 1.44 ± 0.08 , 1.41 ± 0.05 and 1.70 ± 0.03 for Haapiti, Tiahura and Entre 2 Baies respectively. Our model suggests values for these locations of 1.75 ± 0.50 , 1.68 ± 0.47 and 1.70 ± 0.60 respectively. Thus, even according the huge range of the uncertainties, our results highlight the potential to use statistical modeling to predict structural complexity when this information is lacking. Given the importance of structural complexity in ecological functioning

of coral reefs, the reconstruction of structural complexity is critical from long-term benthic historical data, if we want to better understand and predict changes in coral reefs. For example, Graham and Nash (2013) reviewed 20 studies using chain method to measure coral reef rugosity in the Caribbean and found a strong negative relationship between structural complexity and algal cover, a positive relationship between the structural complexity and the coral cover, and a strong positive relationship between structural complexity and fish density and biomass. Later, Graham et al. (2015) demonstrated that structural complexity is the main predictor of coral reef recovery capacity after acute disturbance. This metric represents thus a key variable of coral reef status, and apodictically exists also regarding the present day flattening of coral reefs under the influence of climate change and human impacts. Back-calculating structural complexity, with due caution, may be important to infer present coral reef status compared to historical or quasi-pristine conditions.

Here, we have documented major changes in benthic assemblages across Moorea's coral reefs (Berumen and Pratchett 2006; Adjeroud et al. 2018). Acropora spp. and Pocillopora spp. were dominant species in 2003 (Berumen and Pratchett 2006) and this was still the case in 2004. Pocillopora spp. then, Acropora spp. were affected by both COTS outbreak and the Oli cyclone more than any other taxon (Kayal et al. 2012). Branching and table-shaped species belonging to the genus Acropora were affected first and most heavily. Then, it was followed by those of sub-branching Pocillopora. Finally, Populations of encrusting Montipora, massive Porites, and other hard-coral assemblages also declined, showing a synchronized collapse with the entire coral communities. From 2011 onward, benthic assemblages started to recover mainly thanks to encrusting coral forms and Pocillopora spp. that is presently the dominating coral form in Moorea. The current Pocillopora-dominated state may be a transitional phase, indicative of either continuing degradation or recovery (Aronson et al. 2004). Our back-calculated structural complexity was able to capture major changes due to COTS outbreak and Cyclone Oli, thus attesting the potential to use statistical modeling when the rugosity has not been empirically

collected. However, the status of coral reefs in 2004 when Acropora spp. was on average ≈10% and the present status (average Acropora cover ≈2%) was not enough to distinguish difference in back-calculated structural complexity. There are two proposed explanations for this observation: (a) our model is calibrated with present-day data. Only two transects in 2017 had assemblages with an Acropora cover higher than 5% which is a consequent statistic assumption according to the Acropora cover in 2004 (9.76 \pm 5.61% in 2004 vs 2.53 \pm 1.90% in 2017). This again results in an extremely cautious interpretation of the results due to a possible underestimation of Acropora spp. in the past years (Aronson and Precht 2006; Alvarez-Filip et al. 2009, 2011). And/or (b) Acropora and Pocillopora taxa harbored branching form and similarly contribute to the complexity (Reichert et al. 2017). Thus, the complexity could be rebuilt according to the coral cover underlying the different coral morphologies (Halford et al. 2004; Graham and Nash 2013; Richardson et al. 2017). In order to sharp our hindcasting, the reproducibility of this method could allow us to find a new area with the needed information (i.e. past Acropora and Pocillopora cover vs post Acropora and Pocillopora cover and past and post rugosity) and to test our model. Waiting for this improvement, the CC could be enough accurate to rebuild the past and to predict the complexity for the coming years. Indeed, today more than ever, global coral reefs are witnessing the effects of climate changes, local impacts and natural stressors. Coral bleaching is affecting global coral reefs with an unprecedented frequency and intensity and the future structural complexity of coral reefs is expected to be reduced by these repeated perturbations (Hughes et al. 2018; Lough et al. 2018). As a consequence a loss of ecological diversity productivity is likely (Alvarez-Filip et al. 2009).

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333	<u>Conflict of interest</u> : On behalf of all authors, the corresponding author states that there
334	is no conflict of interest.

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Figures

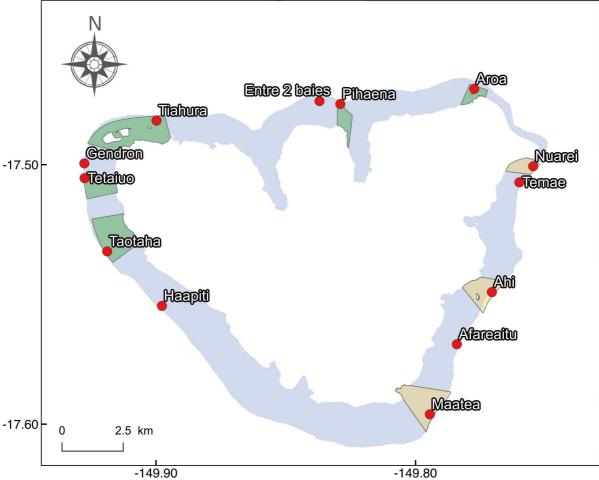


Fig. 1 Location of the 13 sites sampled each year since 2004 from 2017 of the MPA monitoring (red dots) around the island of Moorea. The 5 fully protected MPAs are highlighted in green and the restricted MPAs are highlighted in yellow. The 5 sites outside of the MPAs are controls.

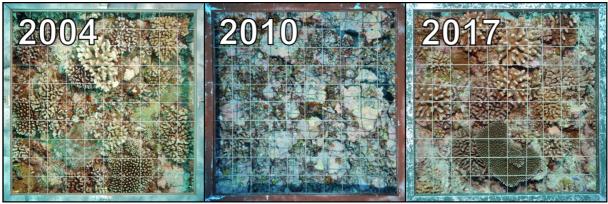


Fig. 2 Evolution of one of the twenty quadrats used to define the coral cover in Haapiti (South West of the island) before (in 2004), during (in 2010) and after (in 2017) Cyclone Oli.

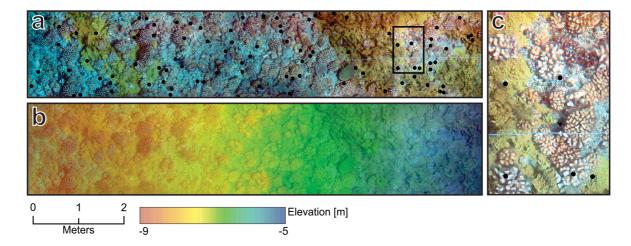


Fig. 3 Example of results from underwater photogrammetry. a) Orthorectified photomosaic. The black dots indicate the random points where the shape classification has been carried out. b) Digital Elevation Model representing depth values (the photomosaic is kept in transparency in the background). c) Detail of the photomosaic.

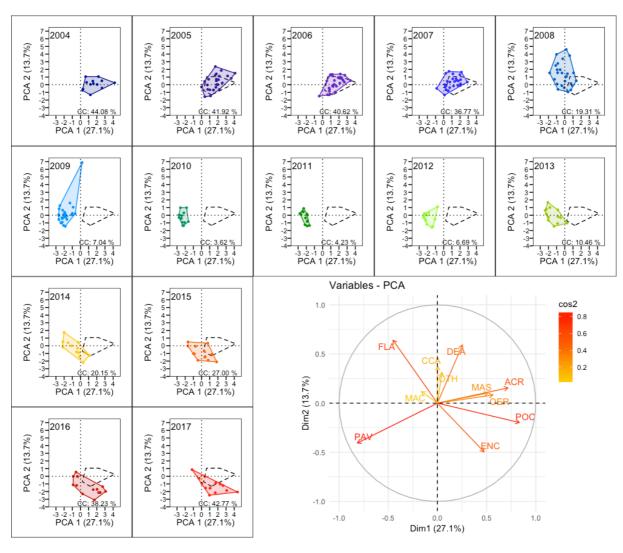


Fig. 4 Principal Components Analysis (PCA) using the 11 morpho-species respecting the code as follows: ACR - Acropora spp; CCA - Crustose Coralline Algae; DEA - Dead Coral; ENC - Encrusting Corals; FLA - Flat (Sand or Mud); MAC - Macroalgae; MAS - Massive corals; OER

- Other Erects Forms; OTH - Others (Sponges or benthic species); PAV - Pavement; and POC - Pocillopora spp. The PCA was used year by year from 2004 to 2017 and the coral cover (CC) is written at the bottom right of each box. The coral community in 2004 is referred as model in black dashed lines each year.

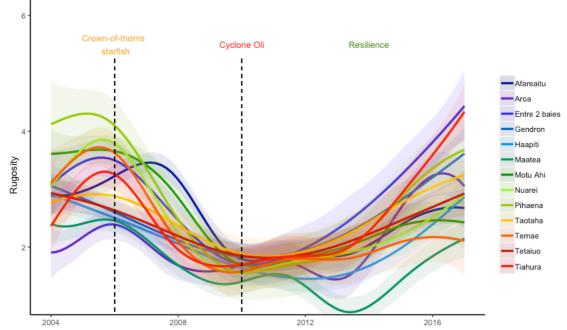


Fig. 5 Rugosity reconstruction from 2004 to 2017 according to the average model and according to the 13 sites around the island of Moorea. Both biologic invasion and extreme climatic weather events are shown for respectively 2006 and 2010.

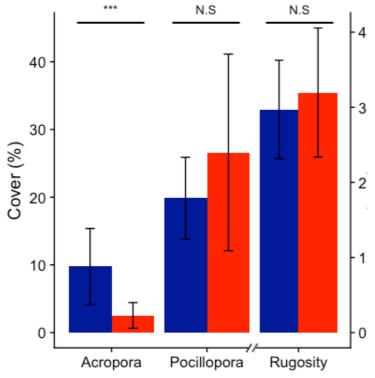


Fig. 6 Difference between the Acropora and the Pocillopora cover (%) on the left, and the difference in rugosity (index) on the right. The dark blue color represents 2004 instead of 2017 is representing in red. The p-value is represented on the top of each barplot according to the

significant R code (*** highly significant (<0.001), **very significant (<0.01), * significant (<0.05), . almost significant (<0.1), N.S non-significant; threshold: p-value = 0.05)

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Tab. 1 Categories of shape classification defined and used to rebuild the rugosity these last 14 years. 9 variables are morphologic instead a distinction at the genus level is done for Acropora spp. and Pocillopora spp. The CCA was differentiated from the pavement according to their extension: when this later was higher than 100 cm², which corresponds to a projected surface of a circle of radius of 5-6 cm it was considered as CCA rather than pavement. The dead corals category understands rubbles and cobbles. The flattening category represents sand or mud substrate. The categories Encrusting, Other Erect Forms and Massive represent different coral morphologies. Finally, the others category represents mostly benthic organism like echinoid or even sponges.

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ACR	Acropora	spp.
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CCA Coralline Crustose Algae

DEA Dead corals

ENC Encrusting corals

FLA Flat (Mud, Sand)

MAC Macroalgae

MAS Massive corals

OER Corals with other erects forms

OTH Other (like echinoid)

PAV Pavement

POC Pocillopora spp.

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Tab. 2 Coefficients and standard error for each parameter according to best model defined: **Rugosity** ~ **Dim 1** + **Dim 3** + **Dim 5** (AIC = 71.776 and $R^2 = 0.81 \pm 0.12$). The p-value represents the significance of each parameters according to the R code (*** highly significant (<0.001), **very significant (<0.01), * significant (<0.05), . almost significant (<0.1), N.S non-significant; threshold: p-value = 0.05)

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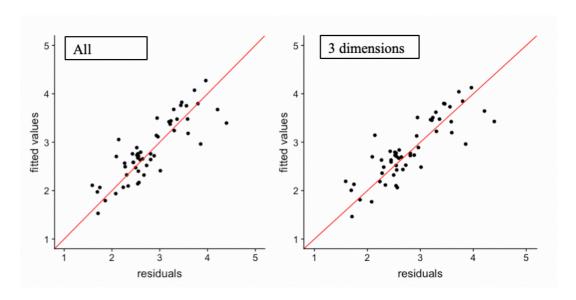
	Estimate	Standard Error	p-value
Intercept	1.65970	0.14616	1.08e-15 (***)
Dimension 1	0.53283	0.08602	9.48e-08 (***)
Dimension 3	0.41917	0.07003	2.03e-07 (***)
Dimension 5	-0.15003	0.06090	1.71e-02 (*)

Appendix

Tab. S1 Post Hoc (Tukey HSD) matrix for testing the rugosity difference according to each year combination. The P-value (threshold: 0.05) for each combination is written in the matrix data as follows. The red values are significantly differents and the blue values are not.

	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
2004	1,00													
2005	0.98	1.00												
2006	0.96	1.00	1.00											
2007	1.00	0.87	0.80	1.00										
2008	0.04	0.00	0.00	0.00	1.00									
2009	0.00	0.00	0.00	0.00	0.93	1.00								
2010	0.00	0.00	0.00	0.00	0.96	1.00	1.00							
2011	0.00	0.00	0.00	0.00	0.95	1.00	1.00	1.00						
2012	0.00	0.00	0.00	0.00	0.98	1.00	1.00	1.00	1.00					
2013	0.01	0.00	0.00	0.00	1.00	1.00	1.00	1.00	1.00	1.00				
2014	0.11	0.00	0.00	0.03	1.00	1.00	0.99	0.99	1.00	1.00	1.00			
2015	0.91	0.05	0.04	0.81	1.00	0.20	0.31	0.30	0.35	0.51	0.98	1.00		
2016	1.00	1.00	0.99	1.00	0.01	0.00	0.00	0.00	0.00	0.00	0.05	0.80	1.00	
2017	1.00	1.00	1.00	1.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.34	1.00	1.00

Fig. S1 Analysis of the residuals (fitted values vs observations) from the total model: Rugosity \sim Dim 1 + Dim 2 + Dim 3 + Dim 4 + Dim 5 (AIC = 71.8, $R^2 = 0.78 \pm 0.08$) on the left and the best model Rugosity \sim Dim 1 + Dim 3 + Dim 5 (AIC = 73.6, $R^2 = 0.81 \pm 0.12$) on the right.



Annex

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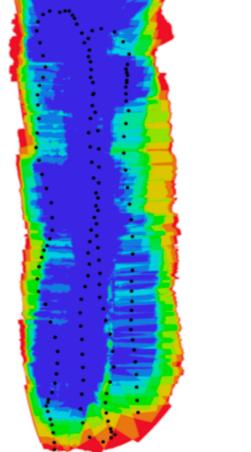
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Annex S1 Report from Agisoft Photoscan for one site, detailing the settings used for the processing of underwater photos.

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1. Survey Data



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2 m

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Fig. 1 Camera locations and image overlap

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Number of images:	157	Camera stations:	157
Flying attitude:	2.56m	Tie points:	139.838
Ground resolution:	0.53 mm/pix	Projections:	383.598
Coverage area:	56.3 sq m	Reprojection error:	1.88 pix

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Tab. 1 Cameras

Camera Model	Resolution	Focal Length	Pixel Size
HERO4 Black (3 mm)	4000 x 3000	3mm	1.73 x 1.73 um

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2. Camera calibration

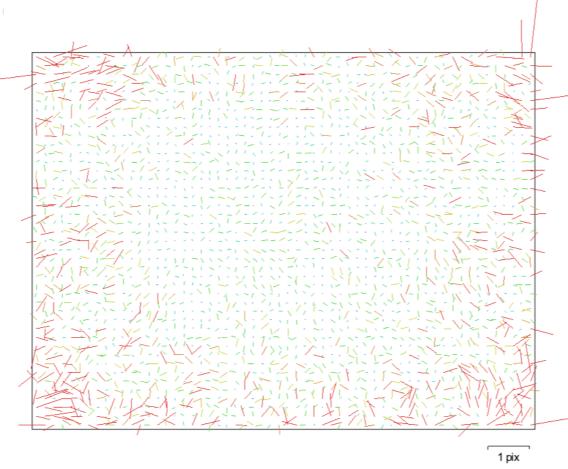


Fig. 2 Image residuals for HERO4 Black (3 mm).

HERO4 Black (3mm) 157 images

Resolution 4000x3000	Focal length 3 mm	Pixel size 1.73 x 1.73 μm	Precalibrated No
Type:	Frame	Skew:	0
Fx:	4666.42	Cx:	2005.25
Fy:	4666.42	Cy:	1486.48
K1:	0.223613	Pĺ:	0.00194777
K2:	0.373779	P2:	-0.00200162
K3:	1.24196	P3:	0
K4:	0	P4:	0

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3. Ground Control Points



Fig. 3 GCP locations

Tab. 2 Control points

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Label	XY error(m)	Z error (m)	Error (m)	Projections	Error (pix)
point1	0.206298	-0.0677987	0.217153	13	1.992
point2	0.0963633	0.0426041	0.105361	16	1.153
point3	0.039409	3.96756e-05	0.039409	17	0.051
point4	0.205045	0.0251539	0.206582	9	0.044
Total	0.154468	0.0419657	0.160067		1.151

2 m

4. <u>Digital Elevation Model</u>

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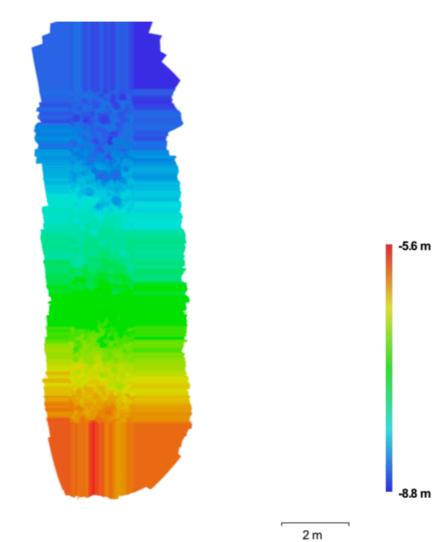


Fig. 4 Reconstructed digital elevation model.

Resolution:

1.06 mm/pix 889956 points per sq m Point density:

599 5. Processing parameters

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General

Cameras 157 Aligned cameras 157 Markers

Coordinate system **Local Coordinates**

Point Cloud

139,838 of 221,231 **Points** RMS reprojection error 0.382302 (1.88392 pix) Max reprojection error 6.30304 (30.1948 pix) 5.32459 pix

Mean key point size Effective overlap 2.94411

Alignment parameters

Accuracy High Pair preselection Disabled Key point limit 40,000 Tie point limit 4,000 Constrain features by mask

Matching time 17 minutes 46 seconds Alignment time 3 minutes 10 seconds

Optimization parameters

Parameters f, cx, cy, k1-k3, p1, p2 5 seconds

Optimization time

Dense Point Cloud

Points 72,229,238

Reconstruction parameters

Quality High Depth filtering Aggressive

Processing time 8 minutes 29 seconds

Model

Faces 4,815,282 Vertices 2,417,898

Reconstruction parameters

Surface type **Arbitrary** Source data Dense Interpolation Enabled Quality High Depth filtering Aggressive Face count 4,815,282

Processing time 4 hours 29 minutes

DEM

Size 1,886 x 9,433 Coordinate system **Local Coordinates**

Reconstruction parameters

Source data Dense cloud Interpolation Enabled

Orthomosaic

Size 3,773 x 18,867 Coordinate system **Local Coordinates**

Channels 3, unit8 Blending mode Mosaic

Reconstruction parameters

Surface Mesh Enable color correction No