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A forgotten element of the blue economy: marine biomimetics and inspiration from the deep sea

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Abstract

The morphology, physiology, and behavior of marine organisms have been a valuable source of inspiration for solving conceptual and design problems. Here, we introduce this rich and rapidly expanding field of marine biomimetics, and identify it as a poorly articulated and often overlooked element of the ocean economy associated with substantial monetary benefits. We showcase innovations across seven broad categories of marine biomimetic design (adhesion, antifouling, armor, buoyancy, movement, sensory, stealth), and use this framing as context for a closer consideration of the increasingly frequent focus on deep-sea life as an inspiration for biomimetic design. We contend that marine biomimetics is not only a “forgotten” sector of the ocean economy, but has the potential to drive appreciation of nonmonetary values, conservation, and stewardship, making it well-aligned with notions of a sustainable blue economy. We note, however, that the highest ambitions for a blue economy are that it not only drives sustainability, but also greater equity and inclusivity, and conclude by articulating challenges and considerations for bringing marine biomimetics onto this trajectory.

Introduction

Nature has been a source of inspiration for humanity throughout history (1, 2). Its influence is evident in the first tools and cave paintings, is reflected in histories, mythologies, and legends, and remains omnipresent today, inspiring everything from the design of airplanes and robots to computer algorithms, packaging, and corporate management structures (3–7). This is the world of biomimicry and biomimetic approaches, which seek to solve conceptual and design problems by mimicking or emulating the structure or performance of organisms and ecosystems that shape the natural world (8–10).

The starting point for biomimetics is the observation of the natural world. Yet the planet's largest habitat—the ocean—has been mostly inaccessible for virtually the entirety of human history. For 3.7 billion years, life has existed in the ocean (three times as long as on land), and the tremendous variety of habitats in the ocean has resulted in comparably diverse morphologies, physiologies, and behavioral mechanisms (8, 11). While it covers 71% of the Earth's surface, the vast majority of the ocean remains infrequently visited (12) and largely unseen. Deep-sea habitats in particular are among the least known on Earth (13). The deepest

point in the ocean, the Challenger Deep in the Mariana Trench, was visited for the first time by humans in 1960—for a total of 20 min—and was not visited again for over 50 y (12, 14, 15).

Ocean exploration in recent decades has resulted in the discovery of deep-sea ecosystems where species have adapted to thrive under extremes of salinity, pressure, light, and temperature, including hydrothermal vents (1979) and brine pools (1983) (16–19). The potential for future discovery is vast: half of the ocean reaches depths of 4,000 m or more, only 20% of the seabed has been mapped (20), and 70% to 90% of marine species remain undescribed (19, 21). Real-time feeds from unmanned deep-sea submersibles outfitted with the latest camera equipment are now freely available (22–24), with thousands of people around the world sharing the experience of seeing unmapped parts of the seafloor and seamounts and unknown species for the first time. Museums around the world are filled with specimens and collections of deep-sea life (25), and thousands of genetic sequences (26), and even complete genomes (27–29), of deep-sea organisms are freely accessible in online databases (30, 31).

Recent advances in ocean science and exploration have occurred alongside a dramatic expansion in the scope and

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diversity of ocean-based activities and industries (32). Today's ocean economy encompasses multiple industries that add up to over USD 1.9 trillion revenues annually (33), including industries as diverse as aquaculture, fisheries, oil and gas, offshore wind, cruise tourism, and marine biotechnology (34). As the ocean economy has grown, so too has attention to issues of equity and inclusivity, as just 100 companies—mostly headquartered in the Global North—accounted for nearly 60% of the ocean economy in 2018 (33, 35). In line with the UN Sustainable Development Goals and Agenda 2030, a variety of aspirational narratives have emerged of the ocean as a source of development that is not only sustainable but also equitable and inclusive. This type of ocean economy has been described by some as a “blue economy,” a term that has become widely used in recent years, but which remains without a broadly agreed definition (35, 36–40) (See note on p. 10). A complementary narrative of “exploration before exploitation” (41) underscores the loss that could accrue from embracing emerging extractive industries in the ocean, such as mining the seabed for metals and minerals, especially in the deep sea where recovery from impacts is extremely slow or impossible (42–44).

If mining and hydrocarbon extraction represent one approach to ocean resources, marine biomimetics represents a starkly different paradigm of resource use: innovation driven by exploration and understanding of the natural world and the life and processes that shape it. Yet in assessments of ocean uses and the blue economy, biomimetics is frequently excluded (33, 34, 45–48), or lumped together as a vaguely defined “emerging industry” (49, 50). We contend that marine biomimetics is a unique element of the ocean economy: vastly diverse and with key benefits for the viability of multiple industries; well-established rather than emerging; and worthy of greater attention in the context of aspirational narratives of a sustainable, inclusive, and equitable blue economy. The diversity of marine species that have spurred innovation also underscores the value of effective ocean conservation, as marine systems grow increasingly stressed by a changing climate and other anthropogenic pressures.

In the following, we first present a review of key focal areas for biomimetic design based on marine life, which then provides context for a closer look at the growing list of instances in which deep-sea life has inspired innovative design and technologies. We conclude by noting benefits that could arise from a more systematic articulation of marine biomimetics in the context of efforts to transform today's ocean economy into a sustainable, inclusive, and equitable blue economy.

Biomimetic design inspired by marine life

The study of marine life has yielded insights into a range of specialized adaptations that allow species to thrive in a diverse range of environmental conditions, and which have been a source of inspiration (8, 51). Categorizing the diversity of ways in which this inspiration translates into applications remains complex, with a wealth of associated terminology (Table 1), and often multiple nonexclusive terms used to describe a single innovation using a variety of available typologies (51–55) (e.g. by function, process, architecture, or material). For instance, the tubercles on humpback whale flippers (see the “Movement” section) have inspired the development of wind turbine blades, which are simultaneously “bio-inspired” (i.e. they are inspired by the structure of a natural material), “biomorphic” (i.e. they resemble the shape of a living thing), “biomimetic” (i.e. they mimic the structure of the flipper), and are also an example of “biomimicry” (i.e. they did not directly inspire the wind turbine, but rather a more sustainable and optimized design of the blades) (8, 56, 57).

In this Review, we follow the spirit of Otto Schmitt, who is credited with coining the term “biomimetics” in the 1950s and argued that biomimetics was simply the “transfer of ideas and analogues from biology to technology” (2, 61). A similar instinct for simplification was articulated by Vincent and co-authors, who suggested that biomimetics could be used synonymously with “biomimesis,” “biomimicry,” “bionics,” “biognosis,” “biologically inspired design,” and similar words and phrases implying copying or adaptation or derivation from biology’ (2). While recognizing the potential for further disambiguation, we opt for this broader framing of biomimetics, and introduce seven broad groupings of marine biomimetic applications (Fig. 1).

Adhesion

The survival of many marine organisms depends on their capacity to adhere to underwater surfaces. The biomineralized adhesives produced by barnacles and oysters as well as the adhesive “byssus threads” used by mussels allow for permanent adhesion, while viscous adhesive proteins secreted by echinoderms like sea stars, sea cucumbers, and sea urchins allow for temporary adhesion, locomotion, and handling of food (10, 86–90). The bioadhesive glue of limpets are up to 97% water, yet are comparable in strength to the cements of oysters and barnacles, and the diversity of marine invertebrate and diatom species that produce bioadhesive gels represent a vast research frontier (86, 88, 91, 92). The biomimetic potential associated with understanding the structure and chemistry of marine bioadhesives has applications across diverse medical fields focused on bone repair (10, 93), dentistry (94, 95), tissue engineering (96), and as surgical sealants (97–99), as well as in the construction of vessels and facilities in the marine environment, particularly when these require coatings and paints that need to adhere to water-facing surfaces (100–104). Similarly, proteins in the byssus threads that mussels use to attach to surfaces have inspired the development of adhesives, which are infused with cerium-oxide nanoparticles (105) to provide anticorrosion properties when applied to metal surfaces (106, 107).

Antifouling

Wherever solid surfaces are found in the ocean, marine organisms begin to adhere to them, using the whole suite of chemical and structural adhesive capacities described in the previous section. This is a process called biofouling, namely the unwanted accumulation of such organisms to everything from submarine hulls to the cooling water intake pipes at nuclear power plants, and the implications for marine industries are severe. Biofouled ships, for instance, consume more fuel due to greater weight and increased friction, and potentially transfer invasive species across oceans (108, 109). The economic losses associated with marine biofouling currently cost marine industries over USD 150 billion annually (108, 110, 111), with estimates of the global market for marine coatings predicted to top USD 15 billion by 2024 (108). Conventional biocides and antifouling paints used for antifouling carry a heavy environmental impact, including bioaccumulation of organotins and copper in marine mammals and other marine life (112, 113). Biomimetic approaches to address these issues have focused in particular on the natural coatings and compounds produced by marine organisms (108, 114, 115), including ascidians (116), macroalgae, (117) algal compounds (118), marine bacteria (119, 120), and sponges, presumably in an effort to avoid biofouling themselves (121). Another rich source of inspiration has been the development of biomimetic surfaces based on the microtopography of marine organisms that function

Table 1. Key concepts associated with nature-inspired activities

Concept	Description	Sustainability	Examples [inspired by]	Reference
Biomimicry	Learning from and then emulating nature's forms, processes, and ecosystems to create more sustainable designs	Main focus	Shinkansen 500 series bullet trains in Japan [kingfisher] (58), ventilation in Eastgate Centre, Harare [termite nest] (59)	Janine Benyus, 1997 (60)
Biomimetics	The process of mimicking the formation, structure or function of a biologically produced substance or material to produce or synthesize an artificial product. Derived from the Greek words <i>bios</i> (life) and <i>mimesis</i> (to imitate), biomimetics was coined by inventor Otto Schmitt (61, 62).	Frequently	Velcro [burdock] (63), gecko tape [gecko] (64), sharkskin swimsuit [sharks] (65)	Otto Schmitt, 1969 (62)
Bionic	The creation of modern engineering systems or a set of functions, based on biological systems and methods found in nature (or using artificial materials and methods to produce movement in a person or animal)	Rarely	Self-healing concrete [multiple organisms] (66, 67), implants in humans [axolotl] (68)	Jack Steele, 1960 (69)
Biophilia/biophilic design	Human tendency to interact or be closely associated with other forms of life in nature: a desire or tendency to commune with nature; biophilic design is an extension of biophilia	Important	Green or living walls inside offices; natural patterns, like curves and fractals, used in interior design [forests, meadows, waterfalls]	E.O. Wilson, 1986 (70)
Biomechanics	Study of the mechanical physics of biological processes or structures	Occasionally	Studying the aerodynamics of bird (71) and insect (72) flight and the hydrodynamics of swimming in fish (73)	Giovanni Alfonso Borelli, 1680 (74)
Bio-utilisation	The direct use of nature for ecological benefits	Occasionally	Gathering medicinal plants; growing algae to make biofuels; cultivation of <i>Artemisia annua</i> to produce malaria drug artemisinin (75)	Youyou Tu, 2011. (75)
Bioremediation	Use of microorganisms, plants, or enzymes, to detoxify contaminants in soil or other environments	Main focus	Using bacteria to break down oilspills (76, 77), or plants to bind, extract, and clean up heavy metals (78, 79)	Vidali, 2001 (80)
Biomorphic	Resembling or suggesting the forms of living organisms, often in design and art	Rarely	The Sagrada Família church by Antoni Gaudí [seashells, trees]; the citrus press "Juicy Salif" by Philippe Starck [squid] (81)	Geoffrey Grigson, 1935 (82)
Bio-affiliation	The idea that humans feel better and are healthier when in contact and connected with nature	Social sustainability	Therapeutic gardens for the elderly; parks in urban areas promoting physical activity [groves, meadows, streams]	Roy Remme et al, 2021. (83)
Bioinspired	General description of several of the concepts in this table, but also a way to emphasise that the resulting idea or innovation is not about simply copying nature	Context dependent	See all above [multiple]	Julian Vincent et al, 2006 (84)
Nature-based solutions	Actions that are inspired and supported by nature, seeking to build resilience while providing social, environmental and economic benefits.	Main focus	Tree-planting, coastal zone restoration [forests, mangroves, coastal dunes]	Alexandre Chausson et al, 2020 (85)

as natural antifoulants (115), including common marine shells, (122) crustaceans (123), seaweed (124), and sharkskin (125, 126).

Armor

The scales of fish have evolved to provide multiple benefits, most notably providing armoured protection without sacrificing flexibility. The scales of fish (and snakes) were likely already inspiring the development of armors in human antiquity, and Ehrlich notes historical examples extending back to the time of the Persian and Roman empires in which scale armor is referenced or depicted (86, 127). The flexibility and protective properties of scales continue to inspire armor designs today (128, 129). The shells of mollusks have also inspired biomimetic designs [including military armor

(130, 131)], with their nacreous layers outperforming conventional ceramics with regard to toughness and both tensile and compressive strength (132). The biomineralization process through which shells form has been the subject of substantial research (133–135), and multilayered ceramics and composites inspired by seashell nacre are in development (132, 136). Mantis shrimps, a frequent source of inspiration (also see the "Sensory" section), possess a pair of appendages called dactyl clubs that they use to strike prey at speeds comparable to a fired bullet with a force of up to 153 kg (although mantis shrimps themselves only weigh 12 to 90 g) (137). The periodic helical structure of the layered fibers in mantis shrimp dactyl clubs and lobster claws have been studied and resulted in bioinspired fiber composite laminates with

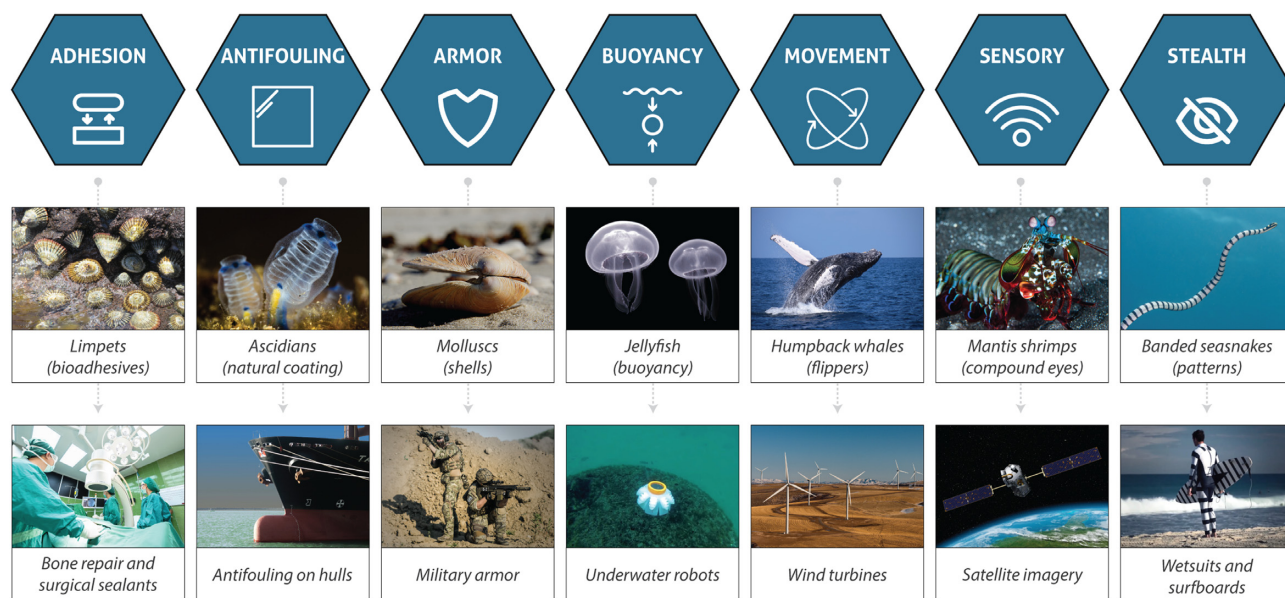


Fig. 1. Marine biomimetics. The diverse morphological, physiological, and behavioral characteristics of marine species have inspired innovations that extend across diverse industries. Photo credits from left to right, first row: NOAA; Christian Gloor; CC0 Public Domain; Luc Viatour; CC0 Public Domain; Cédric Péneau; Christian Gloor. Second row: CC0 Public Domain; CC0 Public Domain; CC0 Public Domain; Jennifer Frame/JenniFish Ocean Test; CC0 Public Domain; NASA; SAMS.

increased resistance to impact force, denting, and cracking (138, 139).

Buoyancy

Neutral buoyancy can be an important asset for movement underwater (140–142), and led to the evolution of swim bladders in fish some 400 million years ago (51). These gas-filled sacs are flexible and adjustable, with fish equilibrating them to provide neutral buoyancy at the topmost reaches of their respective habitats (51, 143). Biomimetic design based on the swim bladder can be found during the Renaissance, with Giovanni Borelli publishing drawings of what could have been the world's first submarine in 1680 (8, 74) (although no evidence exists that this project moved beyond conceptual drawings). Borelli's illustrated plate (Fig. 2) includes not only biomimetic use of goatskin bags as a hydrostatic mechanism to submerge the submarine, but also a conceptualized diver who can move underwater using a large goatskin bag that was meant to double as a source of air and an adjustable buoyancy device (74). The latter innovation foreshadowed a modern-day buoyancy compensation device, that itself mimics the fish's swim bladder, and enables divers to closely control their buoyancy (51). Buoyancy is a ubiquitous concern for operations at sea and underwater, and today there are a number of underwater robots, autonomous gliders and submersibles that have drawn inspiration not just from fish and their buoyancy-related movements (144), but from marine species as diverse as whales (145, 146), dolphins (147, 148), jellyfish (149–151), lobsters (149, 152, 153), and copepods (154).

Movement

The movement of life below water has been perhaps the richest—and one of the most varied—sources of inspiration for biomimetic design. During the Renaissance, Juliana Berners (15th century) and Leonardo da Vinci (16th century) were already remarking on the movement of water eddies, with the latter noting how

the streamlined shape of fish could reduce drag (8, 155, 156). The fusiform design of some fish, with rounded heads and bodies that gradually taper back to the tail, inspired (unsuccessful) biomimetic boat design efforts in the early 19th century based on close studies of the movements of dolphins and trout, and, much later, (successful) design of nuclear submarines (8, 157). While the shape and movement of dolphins continue to be a source of inspiration (147, 158), finned fishes too have drawn intense interest (159–161), as have sharks (162, 163) and rays (164, 165). The fluid mechanics associated with rounded tubercles on the flippers of humpback whales have inspired design both below and above water, most notably in the shape of wind turbines, tidal turbines, and even surfboards (56, 57, 166, 167), while the flexible waving of macroalgae has led to the development of kelp-inspired wave energy generators (168). The body design and propulsive systems of other marine life have inspired additional libraries of biomimetic design, including the jet propulsion and shape of squids (51, 169–171) and other mollusks (172), the movement of siphonophores (173, 174), and the bell shape and contractions of jellyfish (51, 151, 169, 175–177). Applications extend from underwater vehicles all the way to the design of robots and spaceships specially adapted to explore other planets with starkly different atmospheric and gravitational conditions (178). Recent advances in experimental modeling methods have even rendered the shape and movement of extinct marine animals such as plesiosaurs a source of rich biomimetic inspiration for the design of underwater robotics (179–184).

Sensory

The sensory environment under water is distinctly different from above, with water rapidly absorbing light, and currents dispersing chemical trails (51). Consequently, marine life possesses sensory capacities particularly adapted to this environment. Elephant seals, for instance, vibrate their facial whiskers to help locate and pursue prey (185), while dolphins and some whales are able to

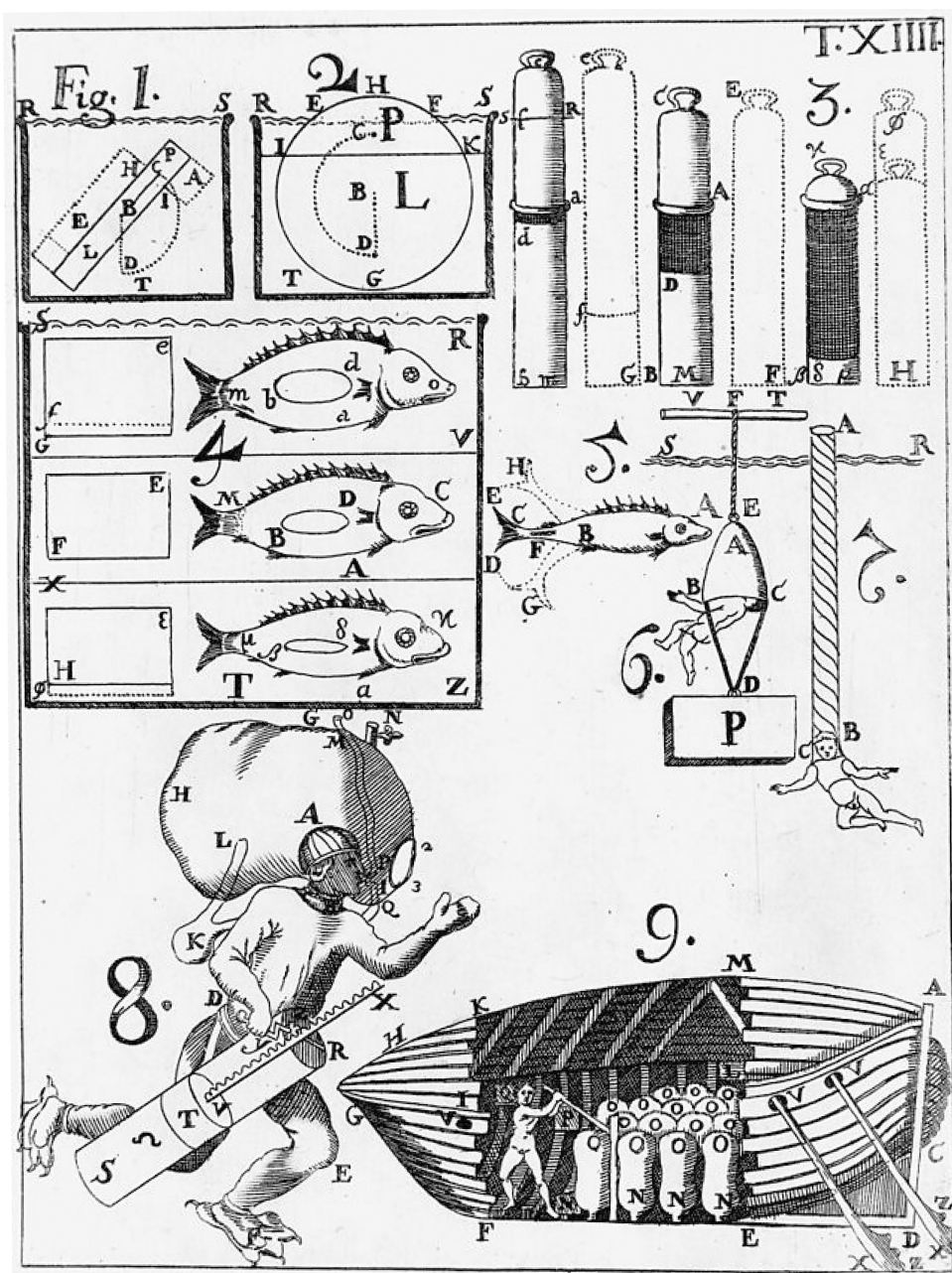


Fig. 2. Panel from “On the Motion of Animals” (*De Motu Animalium*) by Giovanni Alfonso Borelli (1680) illustrating the properties of fish swim bladders (upper left), and how they have inspired biomimetic design of a submarine using air-filled goatskin bags (lower right) as well as a diver’s buoyancy device and tank of air (lower left) (74).

produce high-frequency sounds that enable them to echolocate prey (51, 186, 187). Dolphins and humpback whales also famously use bubble nets to gather their prey into confined spaces, and while traditional sonar fails in environments with heavy sedimentation or bubbles, they still find their prey (51, 188, 189). Biomimetic work on developing dolphin-inspired sonar and radar is ongoing, leading to the development of sonar movement tags for tracking animal movements (190), with vast potential implications for maritime navigation (191). Another novel sensory adaptation is found in sharks, which use their electrosensory system to detect the weak electric field generated by fish. Sharks also possess a “cerebellum chip” that identifies their own electric field and is able to account for this when processing external electromagnetic signals (51). Both the system of electroreceptors and the shark’s ability to process this data have inspired the development

of various innovations including sensors and artificial electroreceptors (51, 192, 193). A final sensory adaptation of note in the marine world is optical, as exemplified perhaps most spectacularly in the world’s 450 species of mantis shrimp. Among other things, mantis shrimp eyes are compound, can move independently of one another, rotate up to 70 degrees in any direction, and see 16 channels of color (humans can process three) as well as ultra-violet and polarized light (194). Associated biomimetic work has focused on adaptively enhancing contrast vision (195), developing novel photodetectors (196), and enhancing satellite imagery (197).

Stealth

Marine life has evolved a diverse range of stealth adaptations, including passive camouflage that mimics background environment, and translucency that renders species almost invisible (51).

Some species seek to mimic the appearance of other species that are poisonous, an approach also used in the design of banded surfboards and wetsuits meant to deter shark attacks by mimicking the patterns of banded seasnakes (51, 198). The active camouflage used by cephalopods like octopi and cuttlefish to opportunistically mimic substrates has drawn intense interest (199) due among other things to its rapidity, with chromatophores that can flash different colors roughly five times every second (51). Cephalopod chromatophore cells have inspired the development of artificial skin (200, 201), paint-like coatings that can be triggered to change color (202, 203), and artificial chromatophores (204, 205).

Other applications

While the previous subsections showcase some of the broader categories of marine biomimetics, they are pieces of a vibrant and rapidly expanding discipline. A comprehensive list would include the many biomimetic designs based on chitin (206–209) and collagen (210–212), design and automation based on the schooling behavior of fish (213, 214), architecture inspired by marine species (215), electrochemical batteries inspired by electric rays (51, 216), use of coral skeletons as bone tissue scaffolds (217–220), and many other innovations. In an even broader sense, groups of organisms and even entire ecosystems can be a source of inspiration, as in the case of coral reefs serving as an inspiration for industrial symbiosis (221) or nature-based solutions (Table 1) such as artificial reefs and coastal protection measures (222, 223).

Looking further to the periphery of marine biomimetics, two additional categories of innovation are of interest. The first of these, marine biodiscovery, often extends beyond biomimetics into the world of bioutilization, with marine natural products (secondary metabolites produced by marine organisms) being collected and subsequently used or modified to produce both medical and nonmedical products. Some 700 new marine natural products are being discovered on an annual basis, and a total of nearly 40,000 have been identified to date (224, 225). In some cases, the proteins associated with marine natural products provide direct inspiration for the development of novel synthetic constructs. Iconic examples of commercial products originating from marine biodiscovery include a suite of marine drugs (20 or which are in clinical use today (226), while at least 33 more are in clinical trials) (227–229). The rates of successful drug discovery from marine natural products is up to four times higher than their terrestrial counterparts, suggesting rich further potential (230, 231). Looking beyond the specific instance of marine drugs, the study of marine life has been crucial for fundamental breakthroughs in medical science and beyond, including for instance the discovery of green fluorescent protein in the jellyfish *Aequorea victoria* (used for broad range of medical and biological applications) (232, 233), and understanding the cell cycle through experiments on sea urchins (and subsequent discovery of cyclins and associated drugs) (234). Collectively, 13 Nobel Prizes in chemistry and medicine have been awarded for work on marine organisms (235).

The second category of innovations located on the periphery of marine biomimetics does not look at individual organisms and ecological systems, but rather the conditions that create their environments, namely geomimetics (material design inspired by natural geological syntheses and natural materials formation) (236). An extreme case under this umbrella is geomimetic design based on hydrothermal synthesis (i.e. methods for crystallization under conditions of high pressure and temperature), although analogue systems exist not only in the ocean

(e.g. hydrothermal vent systems, which can be found in both the shallow and deep sea), but also hot acidic springs on land (236). Hydrothermal synthesis has been used to generate both inorganic materials such as zeolite and synthetic gemstones, as well as organic “green” polymers (237) and polyimide-based covalent organic frameworks, which are promising materials for use as anodes in lithium-ion batteries (236, 238).

The deep sea as a source of inspiration

From an anthropocentric perspective, the deep sea is a place of extremes: the largest biome on Earth, where sunlight does not penetrate, where pressures force us into thick-walled submersibles, and with a diversity of environments characterized by extremes (239). But what is extreme to us is attractive to others, with entire classes of organisms that thrive in extremes of temperature (thermophiles), acidity (acidophiles/alkaliphiles), salinity (halophiles), pressure (barophiles), and high metal concentrations (metalophiles) (236, 240). All such conditions are present in the deep sea, and often in combination, providing habitats for polyextremophiles (microorganisms that benefit from environments characterized by multiple of these “extreme” conditions) (236, 241).

Wherever we have looked for life in the deep ocean, we have found it, with over 25,000 deep-sea species already included in the World Register of Deep-Sea Species (WoRDSS), and likely orders of magnitude more yet to be described (11, 19, 242). While various depths have been suggested as the starting point of the “deep ocean” (243), here a 500 m threshold is considered in line with the WoRDSS database (239, 242). In contrast to the “Biomimetic design inspired by marine life” section, which provides a brief review of broad categories of marine biomimetics, the following subsections focus on individual deep-sea species and species groupings that have been a source of inspiration for diverse applications.

Deep-sea fishes

All the better to see you with: the brownsnout spookfish and glasshead barreleye fish

No light extends beyond 1,000 m into the ocean, but some faint traces of residual daylight persist at depths from 500 to 1,000 m, and a range of species in this zone produce bioluminescence (244) creating a unique sensory environment. A number of mesopelagic teleost fishes have evolved unique eyes that benefit from both sources of light (through a combination of reflective and refractive optics) (Fig. 3A), with the two most-studied species being the brownsnout spookfish (*Dolichopteryx longipes*) and the glasshead barreleye fish (*Rhynchohyalus natalensis*) (244–247). The reflective eyes of decapods like lobsters spurred advances in the field of astrophysics, namely the development of “lobster-eye” X-ray telescopes (248, 249), and the biomimetic design potential of further telescope innovations focused on the unique tilting of mirror plates in *D. longipes* eyes has been suggested (250, 251). A recent study created advanced models to test the optical performance of *D. longipes*, paving the way to the development of optical systems that can function in harsh environments including the deep sea (252). Other examples of biomimetic design are found in architecture, where the mirrored eyes of *D. longipes* inspired concepts of energy-saving roof designs by making optimal use of available daylight (253, 254), as well as in the design of a streamlined passenger car based on the body line of *R. natalensis* (255).



Fig. 3. Inspiration from deep-sea species. Life in the deep sea is specially adapted to thrive in areas characterized by high pressure, absence of sunlight, and limited nutrients. Examples include (A) the mirrored eyes of mesopelagic teleost fish like the glasshead barreleye fish; (B) the transparent teeth of the deep-sea dragonfish; (C) the morphology of the Mariana snailfish (note the CT scan revealing a small crustacean, the green shape, in the snailfish's stomach); (D) antifouling compounds extracted from deep-sea sponges; (E) the air-filled shell that provides buoyancy to the Ram's horn squid; and (F) the armored shell of the Scaly-foot snail. Photo credits: (A) and (B) MBARI; (C) Adam Summers/University of Washington; (D) CoralFISH/Havforskninginstituttet; (E) Schmidt Ocean Institute; (F) Chong Chen.

All the better to eat you with: the deep-sea dragonfish

While the structural properties of marine species have inspired the development of novel ceramics and related materials, such efforts have often focused on the nacreous layers of (coastal) mollusk shells. In the deep sea, the dragonfish (*Aristostomias scintillans*) initially attracted study due to its uniquely transparent teeth, which are thought to be an adaptation enabling further stealth, with the teeth becoming virtually invisible even in proximity to light produced by bioluminescent species (Fig. 3B) (256). A materials science approach to understanding *A. scintillans* teeth found that their transparency arises from a unique nanoscale structure, which also contributes to levels of hardness and sharpness comparable to the teeth of piranhas and great white sharks (256) and a source of inspiration for researchers looking to develop transparent ceramics (257).

Moving under pressure: the Mariana snailfish

The morphology and movement of dolphins and trout have inspired vessel design in shallow waters. Likewise, the development of vessels that can move in the deep sea benefits from close study of deep-sea life. In 2017, scientists discovered a new hadal snailfish species (*Pseudoliparis swirei*) (258) in the Mariana Trench and collected specimens at depths of over 6,000 m (29) (Fig. 3C). A close study of its morphology in 2019 (29) contributed to the spectacular development of an untethered soft robot with multiple points of inspiration from *P. swirei*, including the use of thin, flapping pectoral fins, and the distribution of comparatively heavier electronics within the robot's "head" similar to the distributed weight of the snailfish's skull (259). The soft robot was successfully field-tested at a depth of 10,900 m in the Mariana Trench, underscoring the potential of increased deep-ocean exploration through the

development of additional soft robots that can function in conditions of extreme pressure (259).

High-performance slime: the Atlantic hagfish

The Atlantic hagfish (*Myxine glutinosa*) is an ancient species that is recognizable in fossils from 300 million years ago, and which is unique in the animal kingdom as having a skull but no spinal column (260). The hagfish's defensive slime—which it produces in vast quantities within fractions of a second and is characterized by thousands of silklike protein threads (260)—has attracted curiosity for hundreds of years and transcended the scientific community to enter popular culture (261–263). A search of the World Intellectual Property Organization (WIPO) Patentscope database (264) in July 2022 found 1,170 patents that reference the hagfish, with applications including use of slime threads for high-performance fibers, safety helmets, bulletproof vests, and even antishark sprays (261, 262, 265, 266).

Deep-sea sponges

Skeletons of the deep: deep-sea sponges

Deep-sea sponges have attracted intense interest, as they are known to form siliceous skeletons characterized by high levels of porosity that can act as natural tissue scaffolds (267–270). A challenge of treating bone defects in humans is the need for bone substitution materials, with the traditional sources being the patient themselves (autogenous grafts), other individuals of the same species (allograft materials), or nonhuman species (xenografts) (271). The former is frequently not available, and the latter two carry risk of disease transmission and host rejection, posing a major biomedical challenge (271). Siliceous scaffolds provide an attractive source of either xenografts or biomimetic constructs inspired by their structure (268). A recent study found diversity in porosity and pore size among representative deep-sea sponge species (*Geodia barretti*, *Geodia atlantica*, *Stelletta normani*, *Phakellia ventilabrum*, and *Axinella infundibuliformis*), suggesting varying inspirations for tissue engineering applications (268).

High-definition flow simulations of the deep-sea sponge *Euplectella aspergillum* have shed light on the internal architecture of sponge skeletons and their capacity to reduce hydrodynamic stress (272), while other simulations of its grid-like structure and bracings resulted in biomimetic lattice geometries with implications for the design and resilience of modern infrastructure, and minimizing the amount of material needed for construction (215, 273, 274). One of the most iconic such examples is “The Gherkin” in London's financial district, which due to its design is claimed to function on half the energy of a similarly sized office building (215). Similar structural analysis of the giant anchor spicules of *Monorhaphis chuni* have found optimized designs highly resistant to fractures and cracking, while simultaneously possessing optical properties enabling the transmissibility of visible light with potential fiber-optic applications (275, 276). The porous skeletons of deep-sea sponges have also inspired biomimetic design, with synthetic porous carbon fibers found to achieve high levels of absorption of oils and organic solvents, suggesting future use in bioremediation (277). Likewise the stiffness and toughness of sponge skeletons has inspired the development of novel multilayered pipewalls (278). Patent filings associated with deep-sea sponges include the production of a novel antifouling compound from *Geodia barretti* (Fig. 3D) (279), and a composite material based on the silicated collagen matrix of the glass rope sponge, *Hyalonema sieboldi* (280).

Other deep-sea fauna

Moving rapidly between depths: the Ram's horn squid

While squids do not have internal buoyancy mechanisms comparable to the swim bladders of fish, an unusual variation is found in the Ram's horn squid (*Spirula spirula*), a deep-sea cephalopod mollusk that practices diel vertical migration, generally remaining at depths of 600 to 700 m during the day to avoid predation, and rising at night to depths of 100 to 300 m to feed (51, 281). The “Ram's horn” of *S. spirula* comes from its capacity to manufacture a hollow gas-filled shell that remains almost entirely within its body, and which grows and adds chambers in an expanding spiral form throughout its life (Fig. 3E) (51). While fish swim bladders can equilibrate somewhat to differing depths and pressures (143, 282), this process is slow. The much more rapid diel vertical migration of *S. spirula* is mediated by its rigid shell (although the shells implode at depths of 1,500 m, marking a clear depth limit for the species) (51, 281). Anderson and co-authors note that the biomimetic design of the *Deepsea Challenger* manned submersible, which descended to the ocean's deepest point (10,911 m) in 2012, drew on both the shape and vertical orientation of *S. spirula*, as well as its shell, which it mimicked with a low-density foam of hollow microballoons (15, 51, 283).

The finest protection: the Scaly-foot snail

The scaly-foot snail (*Chrysomallon squamiferum*) is a gastropod mollusk species that was discovered in 1999 and has since been found on three hydrothermal vent systems at depths of over 2,400 m (284). *C. squamiferum* is heavily armored, enabling it to survive the attacks of crabs that have been known to squeeze *C. squamiferum* in their claws for days (Fig. 3F) (285, 286). Similar to other mollusks, *C. squamiferum* has a multilayered shell, but its iron-infused outer layer is unique (derived from the mineral-rich vent fluids) and followed by a thick organic layer and a third stiff mineralized layer, yielding remarkable protection that has attracted the attention of the US Department of Defense, which has funded research to explore its biomimetic potential for armor development (285, 286).

Nontoxic antifouling: *Streptomyces albidoflavus* strains

The bacterium *S. albidoflavus* is ubiquitous, with strains isolated from sources as diverse as a golf course in Korea (287), a mangrove leaf in China (288), soil in Poland (289), and sediments collected at a depth of 5,100 m in the western Pacific Ocean (290). Noting the many novel compounds that have been identified in marine *Streptomyces*, the latter strain (*S. albidoflavus* sp. UST040711-291) was cultured, and a set of five structurally similar compounds were isolated and studied (290, 291). Testing of the *S. albidoflavus* compounds identified functional properties, associated with the 2-furanone ring, that delivered powerful antifouling outcomes, leading the research group to file a corresponding patent on associated nontoxic antifouling derivatives (290, 292).

Revitalizing potentials: cold-water corals

The cold-water coral *Lophelia pertusa* can be found at depths of up to 3,000 m and is extremely slow growing, with radiocarbon dating suggesting it can live for up to 1,000 y, and form reefs that have been dated at over 40,000 y old (293). It is highly vulnerable to deep-sea fishing gears (294) as well as oil exploration and extraction practices (295). Such environmental concerns have led to experimentation with *L. pertusa* to understand the impacts of sedimentation and discard of drill cuttings (296), as well as development of a biomimetic sensor that could monitor the

deep-water sea-floor for impacts from drilling activities, using a system of cameras interspersed with nubbins and polyps of *L. perstusa* (297). Likewise, while deep-sea sponge skeletons have drawn the most active interest in the development of biomimetic tissue scaffolds for biomedical applications, the structure of deep-sea bamboo corals (*Isididae*) has noted potential as a bone implant or substrate for bone revitalization (86, 270, 298).

A “forgotten” ocean economy sector: current challenges and future opportunities

As illustrated in this Review, the field of marine biomimetics is characterized by diversity: in the diversity of innovations it has generated, in the diversity of species that have provided inspiration (from megafauna to microorganisms) and in the diversity of ocean environments that sustain these species from the coastal zone to the deep sea. Furthermore, the economic value of innovations inspired by marine life is substantial (Box 1). Other key components of biomimetics—imagination, imitation, and inspiration—are universal elements of the human experience, suggesting the potential for marine biomimetics as a productive activity across geographies.

Box 1. Examples of revenues and costs associated with key areas of marine biomimetics.

For comparison, estimated annual revenues of key ocean economy sectors include the offshore wind industry (USD 37 billion), cruise tourism (USD 47 billion), container shipping (USD 156 billion), seafood (276 billion), offshore oil and gas (USD 830 billion) (37).

- **Rust.** According to estimates, in Sweden alone, the annual costs associated with corrosion total approximately SEK 90 billion (USD 9.5 billion) (299). The development of biomimetic adhesive coatings, for instance with ceria nanoparticles inspired by mussel byssus threads, aim to cut corrosion rates (see the “Adhesion” section) (106).
- **Biofouling.** The global market for marine coatings is predicted to top USD 15 billion by 2024, and the economic losses associated with marine biofouling currently cost marine industries over USD 150 billion annually (108, 110, 111) (see the “Antifouling” section).
- **Tissue scaffolds.** In the United States alone, over 49 tissue-engineering companies have been established (including 21 companies in the commercial phase of development and generating sales of an estimated USD 9 billion annually in 2017 (300) (see the “Other application” and “Deep-sea sponges” sections).
- **Robotics.** The global underwater robotics market size is expected to reach USD 6.74 billion by 2025 (301), benefiting heavily from biomimetic design focused on the morphology and physiology of marine species (see the “Armor,” “Buoyancy,” “Movement,” “Deep-sea fishes,” and “Other deep-sea fauna” sections).
- **Pharmaceuticals.** Revenues from five “marine drugs” (FDA-approved pharmaceuticals with compounds derived, synthesized or inspired by naturally occurring marine natural products—Adcetris, Halaven, Lovaza, Prialt, and Yondelis) totaled over USD 12.1 billion from 2011 to 2020 (see the “Other applications” section and Supplementary Material).

Where diversity is less evident, however, is in the beneficiaries of marine biomimetics. The majority of innovations identified in this Review are associated with industries disproportionately headquartered in the world's most highly industrialized countries: shipping, robotics, biomedical/biotechnology, wind energy, etc (33, 36). Similarly, the capacity to cover costs of developing innovations and to subsequently access relevant industry counterparts may be a particular challenge to ensuring these activities are inclusive and the benefits are equitably shared [e.g. the timeline for bringing a new pharmaceutical to market can stretch across decades with costs of up to USD 1 billion (26, 230)]. Although capacity building is a recognized priority of the UN Decade of Ocean Science for Sustainable Development 2021–2030 including for deep-sea research (302, 303), investments in marine science as well as contributions to the peer-reviewed literature remain highly skewed towards the world's most highly industrialized countries (304).

Marine biomimetics has also been largely neglected in blue economy strategies and framings (33, 34, 49, 45–48, 50), and we suggest this may be related to the complexity and diversity of marine biomimetic applications as well as challenges in credibly valuing its economic benefits. While the economic value of innovations inspired by marine life can be estimated in some cases (Box 1), relying solely on monetary valuations would be a missed opportunity for articulating the unique position marine biomimetics could play in the blue economy, and for understanding the full value of nature (40, 305). Marine biomimetics does not depend on continuous marine resource use or extraction, and often results in innovations that reduce pollution, energy loss, or emissions, giving it a vanishingly light footprint alongside the stamping footprints of conventional ocean industries like cruise tourism or offshore oil and gas extraction. This will position marine biomimetics as a useful illustration of a blue economy sector if it can ensure that it not only drives sustainability, but also equity and inclusivity, while also spurring greater support for funding basic research on the ocean and the life it contains.

Marine biomimetics is particularly reliant on access to well-functioning marine ecosystems and the capacity to study these. The degradation of marine ecosystems has been widely documented and reported (46, 306). Iconic marine ecosystems like tropical reefs, for instance, face an existential threat from climate change and increasingly frequent marine heatwaves (307), which are also a risk to deep-sea ecosystems (308). Similarly, some of the most remote and least-studied deep-sea environments on Earth have an uncertain future as proponents of mining the international seabed push towards commercial operations, as hydrocarbon extraction moves into deeper and riskier waters, and with continuing use of destructive fishing gears that persist even in purportedly protected areas (36, 42, 309, 310).

Can articulating the role and potential of marine biomimetics in the ocean economy prompt greater awareness of the benefits derived from the ocean? Can marine biomimetics be a sustainable ocean sector at the very heart of the blue economy, sparking inclusive and equitable collaborations across geographies, cultures, and disciplines? Can marine biomimetics become a clear illustration of the importance of nonmonetary valuation, spurring the protection of marine ecosystems as permanent repositories of inspiration? Can it inspire care, and ultimately a sense of stewardship? In this manuscript, we have illustrated the diverse scope and scale of marine biomimetics, and propose that working to address the challenges and questions in this section represents a rich research agenda and action space for scientists and policymakers alike.

Notes

We define the ocean economy to be the sum of economic activities related to the ocean and its coasts. Some consider the blue economy to be largely synonymous with the ocean economy⁴⁹, while others consider it to implicitly encompass additional dimensions such as sustainability³¹¹ and stewardship³¹². In this paper, we use this term to describe an aspirational vision of an ocean economy that is equitable, sustainable and inclusive, in line with articulations introduced around the 2012 UN Convention on Sustainable Development (Rio+20) emphasizing more equitable sharing of benefits^{313–315} and the Sustainable Blue Economy Finance Principles, which define a sustainable blue economy as one that “provides social and economic benefits for current and future generations; restores, protects and maintains diverse, productive and resilient ecosystems; and is based on clean technologies, renewable energy and circular material flows”³¹⁶.

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Supplementary Material

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Authors' Contributions

R.B.: designed research; analyzed data; wrote the paper. J.-B.J., D.A., F.M., J.C., P.J., A.P., C.W., and H.Ö.: wrote the paper.

References

1. Moberg F. 2021. Den uppfinningsrika planeten. Stockholm(Sweden): Natur & Kultur. 292.
2. Vincent JFV, Bogatyreva OA, Bogatyrev NR, Bowyer A, Pahl A-K. 2006. Biomimetics: its practice and theory. *J R Soc. Interface*. 3:471–482.
3. Gao Z, Shi Q, Fukuda T, Li C, Huang Q. 2019. An overview of biomimetic robots with animal behaviors. *Neurocomputing*. 332:339–350.
4. Tian F, Decker EA, Goddard JM. 2013. Controlling lipid oxidation via a biomimetic iron chelating active packaging material. *J Agric Food Chem*. 61:12397–12404.
5. Halder U, Dey B. 2015. Biomimetic algorithms for coordinated motion: theory and implementation. In 2015 IEEE International Conference on Robotics and Automation (ICRA); Seattle, WA. pp. 5426–5432.
6. Guerrero JE, Maestro D, Bottaro A. 2012. Biomimetic spiroid winglets for lift and drag control. *Comptes Rendus Mécanique*. 340:67–80.
7. Tazzi F, Rossi C. 2014. Biomimicry in organizations: drawing inspiration from nature to find new efficient, effective and sustainable ways of managing business. South Carolina(USA): CreateSpace Independent Publishing Platform.
8. Fish FE, Kocak DM. 2011. Biomimetics and marine technology: an introduction. *Mar Technol Soc J*. 45:8–13.
9. Taubes G. 2000. Biologists and engineers create a new generation of robots that imitate life. *Science*. 288:80–83.
10. Scott AR. 2015. Polymers: secrets from the deep sea. *Nature*. 519:S12–S13.
11. Blasiak R, et al. 2020. The ocean genome and future prospects for conservation and equity. *Nat Sustain*. 3:588–596.
12. FAO. 2022. Fisheries and aquaculture—fisheries and aquaculture—deep-sea ecosystems. [accessed 2022 Sep 29]. <https://www.fao.org/fishery/en/topic/166310/en>.
13. Levin LA, et al. 2019. Global observing needs in the deep ocean. *Front Mar Sci*. 6:241.
14. Greenaway SF, Sullivan KD, Umfress SH, Beittel AB, Wagner KD. 2021. Revised depth of the challenger deep from submersible transects; including a general method for precise, pressure-derived depths in the ocean. *Deep Sea Res Part I*. 178:103644.
15. Than K. 2012. James cameron completes record-breaking Mariana Trench dive. National Geographic. [accessed 2022 Sep 29]. <https://www.nationalgeographic.com/adventure/article/120325-james-cameron-mariana-trench-challenger-deepest-returns-science-sub>.
16. Levin LA, et al. 2016. Hydrothermal vents and methane seeps: rethinking the sphere of influence. *Front Mar Sci*. 3:72.
17. Corliss JB, et al. 1979. Submarine thermal springs on the Galápagos Rift. *Science*. 203:1073–1083.
18. Paull CK, et al. 1984. Biological communities at the Florida Escarpment Resemble Hydrothermal Vent Taxa. *Science*. 226:965–967.
19. Mora C, Tittensor DP, Adl S, Simpson AGB, Worm B. 2011. How many species are there on earth and in the ocean? *PLoS Biol*. 9:e1001127.
20. GEBCO. 2022. Mapping progress. The Nippon Foundation-GEBCO Seabed 2030 Project. [accessed 2022 Sep 29]. <https://seabed2030.org/mapping-progress>.
21. Appeltans W, et al. 2012. The magnitude of global marine species diversity. *Curr Biol*. 22:2189–2202.
22. Ocean Exploration Trust. 2022. Nautilus Live | Ocean Exploration Trust. [accessed 2022 Sep 29]. <https://nautiluslive.org/>.
23. NOAA. 2022. NOAA ocean exploration livestream: camera 1. [accessed 2022 Sep 29]. <https://oceanexplorer.noaa.gov/livestreams/welcome.html>.
24. Schmidt Ocean Institute. 2022. Live from R/V Falkor. [accessed 2022 Sep 29]. <https://schmidtocian.org/technology/live-from-rv-falkor/>.
25. Collins JE, et al. 2021. Strengthening the global network for sharing of marine biological collections: recommendations for a new agreement for biodiversity beyond national jurisdiction. *ICES J Mar Sci*. 78:305–314.
26. Blasiak R, Jouffray J-B, Wabnitz CC, Sundström E, Österblom H. 2018. Corporate control and global governance of marine genetic resources. *Sci Adv*. 4:eaar5237.
27. Scott KM, et al. 2006. The genome of deep-sea vent chemolithoautotroph thiomicrospira crunigena XCL-2. *PLoS Biol*. 4:e383.
28. Nakagawa S, et al. 2007. Deep-sea vent ϵ -proteobacterial genomes provide insights into emergence of pathogens. *Proc Natl Acad Sci*. 104:12146–12150.
29. Wang K, et al. 2019. Morphology and genome of a snailfish from the Mariana Trench provide insights into deep-sea adaptation. *Nat Ecol Evol*. 3:823–833.
30. NCBI. 2022. Genome List—Genome—NCBI. [accessed 2022 Sep 29]. <https://www.ncbi.nlm.nih.gov/genome/browse#!/overview/>.

31. NCBI. 2020. Sequence read archive. [accessed 2022 Sep 29]. <https://trace.ncbi.nlm.nih.gov/Traces/sra/sra.cgi?>
32. Jouffray J-B, Blasiak R, Norström AV, Österblom H, Nyström M. 2020. The blue acceleration: the trajectory of human expansion into the ocean. *One Earth*. 2:43–54.
33. Virdin J, et al. 2021. The ocean 100: transnational corporations in the ocean economy. *Sci Adv*. 7:eabc8041.
34. Jouffray J-B, Blasiak R, Norström AV, Österblom H, Nyström M. 2020. The blue acceleration: the trajectory of human expansion into the ocean. *One Earth*. 2:43–54.
35. Österblom H, et al. 2020. Towards ocean equity. Washington, DC: World Resources Institute.
36. Jouffray J-B, et al. 2021. Blue Acceleration: an ocean of risks and opportunities. Ocean Risk and Resilience Action Alliance (OR-RAA) Report. Stockholm: Ocean Risk and Resilience Action Alliance.
37. Virdin J, et al. 2021. The ocean 100: transnational corporations in the ocean economy. *Sci Adv*. 7:eabc8041.
38. Lee K-H, Noh J, Khim JS. 2020. The blue economy and the united nations' sustainable development goals: challenges and opportunities. *Environ Int*. 137:105528.
39. Cisneros-Montemayor AM, et al. 2021. Enabling conditions for an equitable and sustainable blue economy. *Nature*. 591:396–401.
40. Claudet J, Amon DJ, Blasiak R. 2021. Transformational opportunities for an equitable ocean commons. *Proc Natl Acad Sci*. 118:e2117033118.
41. Cordes EE, Levin LA. 2018. Exploration before exploitation. *Science*. 359:719–719.
42. Amon DJ, et al. 2022. Assessment of scientific gaps related to the effective environmental management of deep-seabed mining. *Mar Policy*. 138:105006.
43. Smith CR, et al. 2020. Deep-sea misconceptions cause underestimation of seabed-mining impacts. *Trends Ecol Evol*. 35:853–857.
44. Levin LA, Amon DJ, Lily H. 2020. Challenges to the sustainability of deep-seabed mining. *Nat Sustain*. 3:784–794.
45. Crona B, et al. 2021. Sharing the seas: a review and analysis of ocean sector interactions. *Environ Res Lett*. 16:063005.
46. United Nations. 2016. The First Global Integrated Marine Assessment: World Ocean Assessment I. vol. 1. New York, USA: United Nations.
47. United Nations 2021 The Second World Ocean Assessment: World Ocean Assessment II. vol. 2. New York, USA: United Nations.
48. DNV. 2021. Ocean's future to 2050: a sectoral and regional forecast of the blue economy. [accessed 2022 Sep 29]. <https://www.dnv.com/oceansfuture/index.html>.
49. European Commission. 2021. The EU Blue Economy Report 2021. [accessed 2022 Sep 29]. <https://op.europa.eu/en/publication-detail/-/publication/0b0c5bfd-c737-11eb-a925-01aa75ed71a1>.
50. OECD. 2016. The Ocean Economy in 2030. [accessed 2022 Sep 29]. <https://www.oecd.org/environment/the-ocean-economy-in-2030-9789264251724-en.htm>.
51. Anderson I, Vincent J, Montgomery J. 2019. Ocean innovation: biomimetics beneath the waves. Boca Raton, FL: CRC Press.
52. Vattam SS, Helms ME, Goel AK. 2010. A content account of creative analogies in biologically inspired design. *AI EDAM*. 24:467–481.
53. Nagel JKS, Schmidt L, Born W. 2018. Establishing analogy categories for bio-inspired design. *Designs*. 2:47.
54. Nagel JKS, Schmidt L, Born W. 2016. Fostering diverse analogical transfer in bio-inspired design. In Proceedings of the ASME 2015 International Design Engineering Technical Conferences & Computers and Information in Engineering Conference; Boston, MA.
55. Perera AS, Coppens M-O. 2019. Re-designing materials for biomedical applications: from biomimicry to nature-inspired chemical engineering. *Philos Trans R Soc Math Phys Eng Sci*. 377:20180268.
56. Fish FE. 2020. Biomimetics and the application of the leading-edge tubercles of the humpback whale flipper. In New D. T. H., Ng B. F., editors. Flow control through bio-inspired leading-edge tubercles: morphology, aerodynamics, hydrodynamics and applications. Cham: Springer International Publishing. pp. 1–39.
57. Shi W, Atlar M, Norman R. 2018. Learning from humpback whales for improving the energy capturing performance of tidal turbine blades. In Ölçer AI, Kitada M, Dalaklis D, Ballini F., editors. Trends and challenges in maritime energy management. Cham: Springer International Publishing. pp. 479–497.
58. Cohen YH, Reich Y. 2016. The biomimicry design process: characteristics, stages and main challenge. In Helfman Cohen Y, Reich Y, editors. Biomimetic design method for innovation and sustainability. Cham: Springer International Publishing. pp. 19–29.
59. Atkinson J. 1995. Emulating the termite. *Zimbabwean Rev*. 1:16–19.
60. Benyus JM, 1997. Biomimicry: innovation inspired by nature. New York, USA: Morrow.
61. Harkness JM. 2002. In appreciation of a lifetime of connections: otto herbert schmitt, 1913–1998. *Phys Perspect*. 4:456–490.
62. Schmitt OH, 1969. Some interesting and useful biomimetic transforms. Proceedings of the Third International Biophysics Conference. Boston, USA.
63. Budde R. 1995. The story of Velcro. *Phys World*. 8:22–22.
64. Ge L, Sethi S, Ci L, Ajayan PM, Dhinojwala A. 2007. Carbon nanotube-based synthetic gecko tapes. *Proc Natl Acad Sci*. 104:10792–10795.
65. Das S, Bhowmick M, Chattopadhyay SK, Basak S. 2015. Application of biomimicry in textiles. *Curr Sci*. 109:893–901.
66. Li Q, et al. 2019. A novel bio-inspired bone-mimic self-healing cement paste based on hydroxyapatite formation. *Cem Concr Compos*. 104:103357.
67. Shah KW, Huseien GF. 2020. Biomimetic self-healing cementitious construction materials for smart buildings. *Biomimetics*. 5:47.
68. Chen X, et al. 2015. The axolotl fibula as a model for the induction of regeneration across large segment defects in long bones of the extremities. *PLoS One*. 10:e0130819.
69. Roth RR. 1983. The foundation of bionics. *Perspect Biol Med*. 26:229–242.
70. Wilson E. O. 1900. Biophilia. Cambridge, MA: Harvard University Press.
71. Tobalske BW. 2007. Biomechanics of bird flight. *J Exp Biol*. 210:3135–3146.
72. Sane SP. 2003. The aerodynamics of insect flight. *J Exp Biol*. 206:4191–4208.
73. Drucker EG, Lauder GV. 2002. Experimental hydrodynamics of fish locomotion: functional insights from wake visualization1. *Integr Comp Biol*. 42:243–257.
74. Borelli GA. 1680. De motu animalium. Rome, Italy: Apud Petrum Gosse.
75. Tu Y. 2011. The discovery of artemisinin (qinghaosu) and gifts from Chinese medicine. *Nat Med*. 17:1217–1220.

76. Ron EZ, Rosenberg E. 2014. Enhanced bioremediation of oil spills in the sea. *Curr Opin Biotechnol*. 27:191–194.
77. Prince RC. 1997. Bioremediation of marine oil spills. *Trends Biotechnol*. 15:158–160.
78. Yaashikaa PR, Kumar PS, Jeevanantham S, Saravanan R. 2022. A review on bioremediation approach for heavy metal detoxification and accumulation in plants. *Environ Pollut*. 301: 119035.
79. Ojuederie OB, Babalola OO. 2017. Microbial and plant-assisted bioremediation of heavy metal polluted environments: a review. *Int J Environ Res Public Health*. 14:1504.
80. Vidali M. 2001. Bioremediation. an overview. *Pure Appl Chem*. 73:1163–1172.
81. Orr C, 2019. Philippe Starck's Juicy Salif lemon squeezer: genius design or just a cool fruit squeezer? *DesignStudies*1. [accessed 2022 Sep 29]. <https://medium.com/designstudies1/philippe-s-tarcks-juicy-salif-lemon-squeezer-genius-design-or-just-a-cool-fruit-squeezer-d2b41634407c>.
82. Grigson G. 1935. *The arts to-day*. London: John Lane.
83. Remme RP, et al. 2021. An ecosystem service perspective on urban nature, physical activity, and health. *Proc Natl Acad Sci*. 118:e2018472118.
84. Vincent JFV, Bogatyreva OA, Bogatyrev NR, Bowyer A, Pahl A-K. 2006. Biomimetics: its practice and theory. *J R Soc, Interface*. 3:471–482.
85. Chausson A, et al. 2020. Mapping the effectiveness of nature-based solutions for climate change adaptation. *Global Change Biol*. 26:6134–6155.
86. Ehrlich H. 2019. *Marine biological materials of invertebrate origin*. vol. 13. Cham: Springer International Publishing.
87. Kamino K. 2016. Barnacle underwater attachment. In Smith AM, editor. *Biological adhesives*. Cham: Springer International Publishing. pp. 153–176.
88. Smith AM. 2016. The biochemistry and mechanics of gastropod adhesive gels. In Smith AM, editor. *Biological adhesives*. Cham: Springer International Publishing. pp. 177–192.
89. Flammang P, Demeuldre M, Hennebert E, Santos R. 2016. Adhesive secretions in echinoderms: a review. In Smith AM, editor. *Biological adhesives*. Cham: Springer International Publishing. pp. 193–222.
90. Silverman HG, Roberto FF. 2007. Understanding marine mussel adhesion. *Mar Biotechnol*. 9:661–681.
91. Molino PJ, Chiovitti A, Higgins MJ, Dugdale TM, Wetherbee R. 2016. Diatom adhesives: molecular and mechanical properties. In Smith AM, editor. *Biological adhesives*. Cham: Springer International Publishing. pp. 57–86.
92. Aldred N, Petrone L. 2016. Progress in the study of adhesion by marine invertebrate larvae. In Smith AM, editor. *Biological adhesives*. Cham: Springer International Publishing. pp. 87–105.
93. Lalzawmliana V, et al. 2019. Marine organisms as a source of natural matrix for bone tissue engineering. *Ceram Int*. 45:1469–1481.
94. Green DW, Lai W-F, Jung H-S. 2014. Evolving marine biomimetics for regenerative dentistry. *Mar Drugs*. 12:2877–2912.
95. Şenel S, Aksoy EA, Akca G. 2019. Application of chitosan based scaffolds for drug delivery and tissue engineering in dentistry. In Choi AH, Ben-Nissan B., editors. *Marine-derived biomaterials for tissue engineering applications*. Singapore: Springer. pp. 157–178.
96. Lalzawmliana V, Mukherjee P, Kundu B, Nandi SK. 2019. Clinical application of biomimetic marine-derived materials for tissue engineering. In Choi AH, Ben-Nissan B., editors. *Marine-derived biomaterials for tissue engineering applications*. Singapore: Springer. pp. 329–356.
97. Heher P, Ferguson J, Redl H, Slezak P. 2018. An overview of surgical sealant devices: current approaches and future trends. *Expert Rev Med Devices*. 15:747–755.
98. Gohar F, et al. 2019. Driving medical innovation through interdisciplinarity: unique opportunities and challenges. *Front Med*. 6:35.
99. Qiu L, See AAQ, Steele TWJ, King NKK. 2019. Bioadhesives in neurosurgery: a review. *J Neurosurg*. 133:1928–1938.
100. Kamino K. 2008. Underwater adhesive of marine organisms as the vital link between biological science and material science. *Mar Biotechnol*. 10:111–121.
101. Cholewinski A, Yang F, Zhao B. 2019. Algae-mussel-inspired hydrogel composite glue for underwater bonding. *Mater Horiz*. 6:285–293.
102. Zhao Y, et al. 2017. Bio-inspired reversible underwater adhesive. *Nat Commun*. 8:2218.
103. Heinzmann C, Weder C, Espinosa LM. 2016. Supramolecular polymer adhesives: advanced materials inspired by nature. *Chem Soc Rev*. 45:342–358.
104. Shao H, Stewart RJ. 2010. Biomimetic underwater adhesives with environmentally triggered setting mechanisms. *Adv Mater*. 22:729–733.
105. Xu C, Qu X. 2014. Cerium oxide nanoparticle: a remarkably versatile rare earth nanomaterial for biological applications. *NPG Asia Mater*. 6:e90–e90.
106. Zhang F, Pan J. 2019. Recent development of corrosion protection strategy based on mussel adhesive protein. *Front Mater*. 6:207.
107. Sababi M, et al. 2012. Corrosion inhibiting coating based on cerium oxide and a catecholic polymer. [accessed 2022 Sep 29]. <https://patents.google.com/patent/CA2804826C/en>.
108. Chen L, et al. 2021. Biomimetic surface coatings for marine antifouling: natural antifoulants, synthetic polymers and surface microtopography. *Sci Total Environ*. 766:144469.
109. Hewitt CL, Gollasch S, Minchin D. 2009. The vessel as a vector—biofouling, ballast water and sediments. In Rilov G., Crooks J.A., editors. *Biological invasions in marine ecosystems: ecological, management, and geographic perspectives*. Berlin, Heidelberg: Springer. pp. 117–131.
110. Manolakis I, Azhar U. 2020. Recent advances in mussel-inspired synthetic polymers as marine antifouling coatings. *Coatings*. 10:653.
111. Nurioglu AG, Esteves ACC, With G. 2015. Non-toxic, non-biocide-release antifouling coatings based on molecular structure design for marine applications. *J Mater Chem B*. 3:6547–6570.
112. Amara I, Miled W, Slama RB, Ladhari N. 2018. Antifouling processes and toxicity effects of antifouling paints on marine environment. A review. *Environ Toxicol Pharmacol*. 57:115–130.
113. Bergman K, Ziegler F. 2019. Environmental impacts of alternative antifouling methods and use patterns of leisure boat owners. *Int J Life Cycle Assess*. 24:725–734.
114. Fusetani N. 2011. Antifouling marine natural products. *Nat Prod Rep*. 28:400–410.
115. Ralston E, Swain G. 2011. Can biomimicry and bioinspiration provide solutions for fouling control? *Mar Technol Soc J*. 45:216–227.
116. Levert A, et al. 2021. Antifouling activity of meroterpenes isolated from the ascidian *Aplidium aff. densum*. *Mar Biotechnol*. 23:51–61.

117. Salama AJ, Satheesh S, Balqadi AA. 2018. Antifouling activities of methanolic extracts of three macroalgal species from the Red Sea. *J Appl Phycol*. 30:1943–1953.
118. Saha M, Goecke F, Bhadury P. 2018. Minireview: algal natural compounds and extracts as antifoulants. *J Appl Phycol*. 30:1859–1874.
119. Dobretsov S, et al. 2011. Inhibition of marine biofouling by bacterial quorum sensing inhibitors. *Biofouling*. 27:893–905.
120. Aguila-Ramírez RN, et al. 2014. Antifouling activity of symbiotic bacteria from sponge *Aplysina Gerardogreeni*. *Int Biodeterior Biodegrad*. 90:64–70.
121. Labriere C, Cervin G, Pavia H, Hansen JH, Svenson J. 2021. Structure–activity relationship probing of the natural marine antifoulant baretin. *Mar Biotechnol*. 23:904–916.
122. Bai XQ, et al. 2013. Study on biomimetic preparation of shell surface microstructure for ship antifouling. *Wear*. 306:285–295.
123. Brzozowska AM, et al. 2014. Biomimicking micropatterned surfaces and their effect on marine biofouling. *Langmuir*. 30:9165–9175.
124. Zhao L, et al. 2020. Layer-by-layer-assembled antifouling films with surface microtopography inspired by *Laminaria japonica*. *Appl Surf Sci*. 511:145564.
125. Carman ML, et al. 2006. Engineered antifouling microtopographies—correlating wettability with cell attachment. *Biofouling*. 22:11–21.
126. Wong FYM, Mak MS. 2021. A study to compare the fouling resistance and self-cleaning properties of two-patterned surfaces with an un-patterned control surface. *J Phys Conf Ser*. 2120:012015.
127. Herodotus. 1996. *Herodotus: The Histories*. New York, USA: Penguin Books.
128. Ehrlich H. 2015. Materials design principles of fish scales and armor. In Ehrlich H, editor. *Biological materials of marine origin: vertebrates*. Netherlands: Springer. pp. 237–262.
129. Miranda P, Pajares A, Meyers MA. 2019. Bioinspired composite segmented armour: numerical simulations. *J Mater Res Technol*. 8:1274–1287.
130. Yadav R, Naebe M, Wang X, Kandasubramanian B. 2016. Body armour materials: from steel to contemporary biomimetic systems. *RSC Adv*. 6:115145–115174.
131. Islam MK, Hazell PJ, Escobedo JP, Wang H. 2021. Biomimetic armour design strategies for additive manufacturing: a review. *Mater Des*. 205:109730.
132. Akella K. 2012. Biomimetic designs inspired by seashells: seashells helping engineers design better ceramics. *Resonance*. 17:573–591.
133. Clark MS, et al. 2020. Deciphering mollusc shell production: the roles of genetic mechanisms through to ecology, aquaculture and biomimetics. *Biol Rev*. 95:1812–1837.
134. Morris JP, Wang Y, Backeljau T, Chapelle G. 2016. Biomimetic and bio-inspired uses of mollusc shells. *Mar Geonomics*. 27:85–90.
135. Currey JD. 1999. The design of mineralised hard tissues for their mechanical functions. *J Exp Biol*. 202:3285–3294.
136. Mishra N, Kandasubramanian B. 2018. Biomimetic design of artificial materials inspired by iridescent nacre structure and its growth mechanism. *Polym-Plast Technol Eng*. 57:1592–1606.
137. Weaver JC, et al. 2012. The stomatopod dactyl club: a formidable damage-tolerant biological hammer. *Science*. 336:1275–1280.
138. Han Q, et al. 2022. Impact resistant basalt fiber-reinforced aluminum laminate with Janus helical structures inspired by lobster and mantis shrimp. *Compos Struct*. 291:115551.
139. Behera RP, Le Ferrand H. 2021. Impact-resistant materials inspired by the mantis shrimp's dactyl club. *Matter*. 4:2831–2849.
140. Zhu Y, Liu Y, Wang S, Zhang L, Wang Y. 2021. A bionic flexible-bodied underwater glider with neutral buoyancy. *J Bionic Eng*. 18:1073–1085.
141. Eastman JT. 2020. The buoyancy-based biotope axis of the evolutionary radiation of Antarctic cryonotothenioid fishes. *Polar Biol*. 43:1217–1231.
142. Macdonald A. 2021. Buoyancy at depth. In Macdonald A, editor. *Life at high pressure: in the deep sea and other environments*. Cham: Springer International Publishing. pp. 271–294.
143. Jones FRH. 1951. The swimbladder and the vertical movements of teleostean fishes. *J Exp Biol*. 28:553–566.
144. Tan X. 2011. Autonomous robotic fish as mobile sensor platforms: challenges and potential solutions. *Mar Technol Soc J*. 45:31–40.
145. Yen J. 2011. Sink and swim: clues from nature for aquatic robotics. *Mar Technol Soc J*. 45:16–18.
146. Clarke MR. 1978. Buoyancy control as a function of the spermaceti organ in the sperm whale. *J Mar Biol Assoc U K*. 58: 27–71.
147. Yu J, Tan M. 2020. Control of yaw and pitch maneuvers of a multilink dolphin robot. In Yu J, Tan M, editors. *Motion control of biomimetic swimming robots*. Singapore: Springer. pp. 123–148.
148. Yu J, Tan M. 2020. Leaping control of self-propelled robotic dolphin. In Yu J, Tan M, editors. *Motion control of biomimetic swimming robots*. Singapore: Springer. pp. 149–172.
149. Shi L, Guo S, Mao S, Li M, Asaka K. 2013. Development of a lobster-inspired underwater microrobot. *Int J Adv Rob Syst*. 10:44.
150. Shi L, Guo S, Asaka K. 2011. A novel butterfly-inspired underwater microrobot with pectoral fins. In 2011 IEEE International Conference on Mechatronics and Automation. pp. 853–858. doi:.
151. Joshi K, Villanueva A, Smith C, Priya S. 2011. Modeling of artificial *Aurelia aurita* bell deformation. *Mar Technol Soc J*. 45:165–180.
152. Ayers J, Westphal A, Blustein D. 2011. A conserved neural circuit-based architecture for ambulatory and undulatory biomimetic robots. *Mar Technol Soc J*. 45:147–152.
153. Ayers J, Witting J. 2007. Biomimetic approaches to the control of underwater walking machines. *Philos Transact A Math Phys Eng Sci*. 365:273–295.
154. Pond DW, Tarling GA. 2011. Phase transitions of wax esters adjust buoyancy in diapausing *Calanoides acutus*. *Limnol Oceanogr*. 56:1310–1318.
155. Smith DL, Goodwin RA, Nestler JM. 2014. Relating turbulence and fish habitat: a new approach for management and research. *Rev Fish Sci Aquac*. 22:123–130.
156. Vinci L. 2012. *The notebooks of Leonardo da Vinci*. Rome, Italy: Courier Corporation.
157. Gibbs-Smith CH. 1962. Sir George Cayley's aeronautics 1796–1855. *Br J Hist Sci*. 1: 286–287.
158. Liu J, et al. 2021. Design and analysis of a novel tendon-driven continuum robotic dolphin. *Bioinspiration Biomimetics*. 16:065002.
159. Lauder G, et al. 2011. Robotic models for studying undulatory locomotion in fishes. *Mar Technol Soc J*. 45:41–55.
160. Tangorra J, Gericke T, Lauder G. 2011. Learning from the fins of ray-finned fish for the propulsors of unmanned undersea vehicles. *Mar Technol Soc J*. 45:65–73.

161. Rufo M, Smithers M. 2011. GhostSwimmer™ AUV: applying biomimetics to underwater robotics for achievement of tactical relevance. *Mar Technol Soc J*. 45:24–30.
162. Dong H, Wu Z, Chen D, Tan M, Yu J. 2020. Development of a whale-shark-inspired gliding robotic fish with high maneuverability. *IEEE/ASME Trans Mechatron*. 25:2824–2834.
163. Long J, et al. 2011. Inspired by sharks: a biomimetic skeleton for the flapping, propulsive tail of an aquatic robot. *Mar Technol Soc J*. 45:119–129.
164. Moored K, Dewey P, Leftwich M, Bart-Smith H, Smits A. 2011. Bioinspired propulsion mechanisms based on manta ray locomotion. *Mar Technol Soc J*. 45:110–118.
165. Moored K, Fish F, Kemp T, Bart-Smith H. 2011. Batoid fishes: inspiration for the next generation of underwater robots. *Mar Technol Soc J*. 45:99–109.
166. Fish FE, Weber PW, Murray MM, Howle LE. 2011. The tubercles on humpback whales' flippers: application of bio-inspired technology. *Integr Comp Biol*. 51:203–213.
167. Shormann DE, Panhuis M. 2020. Performance evaluation of humpback whale-inspired shortboard surfing fins based on ocean wave fieldwork. *PLoS One*. 15:e0232035.
168. Wang Y, et al. 2021. Flexible seaweed-like triboelectric nanogenerator as a wave energy harvester powering marine internet of things. *ACS Nano*. 15:15700–15709.
169. Bujard T, Giorgio-Serchi F, Weymouth GD. 2021. A resonant squid-inspired robot unlocks biological propulsive efficiency. *Sci Robot*. 6:eabd2971.
170. Luo Y, Xiao Q, Zhu Q, Pan G. 2020. Pulsed-jet propulsion of a squid-inspired swimmer at high Reynolds number. *Phys Fluids*. 32:111901.
171. Bartol IK, Krueger PS, Stewart WJ, Thompson JT. 2009. Hydrodynamics of pulsed jetting in juvenile and adult brief squid *Lolliguncula brevis*: evidence of multiple jet 'modes' and their implications for propulsive efficiency. *J Exp Biol*. 212:1889–1903.
172. Wang Y, et al. 2020. Development of a biomimetic scallop robot capable of jet propulsion. *Bioinspiration Biomimetics*. 15:036008.
173. Sutherland KR, Gemmell BJ, Colin SP, Costello JH. 2019. Maneuvering performance in the colonial siphonophore, *nanomia bijuga*. *Biomimetics*. 4:62.
174. Sutherland KR, Gemmell BJ, Colin SP, Costello JH. 2019. Propulsive design principles in a multi-jet siphonophore. *J Exp Biol*. 222:jeb198242.
175. Villanueva A, Smith C, Priya S. 2011. A biomimetic robotic jellyfish (Robojelly) actuated by shape memory alloy composite actuators. *Bioinspir Biomim*. 6:036004.
176. Najem J, Sarles SA, Akle B, Leo DJ. 2012. Biomimetic jellyfish-inspired underwater vehicle actuated by ionic polymer metal composite actuators. *Smart Mater Struct*. 21:094026.
177. Nawroth JC, et al. 2012. A tissue-engineered jellyfish with biomimetic propulsion. *Nat Biotechnol*. 30:792–797.
178. Babu Mannam NP, Mahbub Alam Md, Krishnankutty P. 2020. Review of biomimetic flexible flapping foil propulsion systems on different planetary bodies. *Results Eng*. 8:100183.
179. Muscutt LE, et al. 2017. The four-flipper swimming method of plesiosaurs enabled efficient and effective locomotion. *Proc R Soc B Biol Sci*. 284:20170951.
180. Gutarra S, Rahman IA. 2022. The locomotion of extinct secondarily aquatic tetrapods. *Biol Rev*. 97:67–98.
181. Gutarra S, et al. 2019. Effects of body plan evolution on the hydrodynamic drag and energy requirements of swimming in ichthyosaurs. *Proc R Soc B Biol Sci*. 286:20182786.
182. Liu S, et al. 2015. Computer simulations imply forelimb-dominated underwater flight in plesiosaurs. *PLoS Comput Biol*. 11:e1004605.
183. Troelsen PV, Wilkinson DM, Seddighi M, Allanson DR, Falkingham PL. 2019. Functional morphology and hydrodynamics of plesiosaur necks: does size matter? *J Vertebr Paleontol*. 39:e1594850.
184. Long JH, Schumacher J, Livingston N, Kemp M. 2006. Four flippers or two? Tetrapodal swimming with an aquatic robot. *Bioinspir Biomim*. 1:20–29.
185. Adachi T, et al. 2022. Whiskers as hydrodynamic prey sensors in foraging seals. *Proc Natl Acad Sci*. 119:e2119502119.
186. Coombs EJ, Clavel J, Park T, Churchill M, Goswami A. 2020. Wonky whales: the evolution of cranial asymmetry in cetaceans. *BMC Biol*. 18:86.
187. Mccurry MR, et al. 2021. Brain size evolution in whales and dolphins: new data from fossil mysticetes. *Biol J Linn Soc*. 133:990–998.
188. Leighton TG, Chua GH, White PR. 2012. Do dolphins benefit from nonlinear mathematics when processing their sonar returns? *Proc R Soc Math Phys Eng Sci*. 468:3517–3532.
189. Wiley D, et al. 2011. Underwater components of humpback whale bubble-net feeding behaviour. *Behaviour*. 148:575–602.
190. Goulet P, Guinet C, Swift R, Madsen PT, Johnson M. 2019. A miniature biomimetic sonar and movement tag to study the biotic environment and predator-prey interactions in aquatic animals. *Deep Sea Res Part Oceanogr Res Pap*. 148:1–11.
191. University of Southampton. 2022. Research project: do dolphins think nonlinearly?. [accessed 2022 Sep 29]. https://www.southampton.ac.uk/engineering/research/projects/do_dolphins_think_nonlinearly.page#project_overview%0A.
192. Sun J-Y. 2021. Biomimetic approaches with stretchable ionics. Abstract of the research paper of the Korea Polymer Society Conference. 46:42–42.
193. Makarczuk T, et al. 2011. Biomimetic MEMS to assist, enhance, and expand human sensory perceptions: a survey on state-of-the-art developments. In Schmid U., Sánchez-Rojas J.L., Leester-Schaede M., editors. *Smart sensors, actuators, and MEMS V*. vol. 8066. SPIE. pp. 503–517.
194. Tadepalli S, Slocik JM, Gupta MK, Naik RR, Singamaneni S. 2017. Bio-optics and bio-inspired optical materials. *Chem Rev*. 117:12705–12763.
195. Zhong B, Wang X, Gan X, Yang T, Gao J. 2020. A biomimetic model of adaptive contrast vision enhancement from mantis shrimp. *Sensors*. 20:4588.
196. Altaqui A, et al. 2021. Mantis shrimp-inspired organic photodetector for simultaneous hyperspectral and polarimetric imaging. *Sci Adv*. 7:eabe3196.
197. Olivia. 2020. All eyes on the reef. [accessed 2022 Sep 29]. <https://www.science.org.au/curious/earth-environment/all-eyes-reef>.
198. Hart NS, Collin SP. 2015. Sharks senses and shark repellents. *Integr Zool*. 10:38–64.
199. Phan L, et al. 2016. Dynamic materials inspired by cephalopods. *Chem Mater*. 28:6804–6816.
200. Fishman A, Rossiter J, Homer M. 2015. Hiding the squid: patterns in artificial cephalopod skin. *J R Soc, Interface*. 12:20150281.
201. Fishman A, Catsis S, Homer M, Rossiter JM. 2018. Smart squid skin: patterns in networks of artificial chromatophores. In *Proceedings of the Electroactive Polymer Actuators and Devices (EAPAD) XX*. vol. 10594. pp. 379–385. SPIE.

202. Martin CA, et al. 2021. Biomimetic colorants and coatings designed with cephalopod-inspired nanocomposites. *ACS Appl Bio Mater.* 4:507–513.
203. Wang W et al. 2015. Easy approach to assembling a biomimetic color film with tunable structural colors. *J Opt Soc Amer A.* 32:1109–1117.
204. Han D, Wang Y, Yang C, Lee H. 2021. Multimaterial printing for cephalopod-inspired light-responsive artificial chromatophores. *ACS Appl Mater Interfaces.* 13:12735–12745.
205. Rossiter J, Yap B, Conn A. 2012. Biomimetic chromatophores for camouflage and soft active surfaces. *Bioinspir Biomim.* 7:036009.
206. Spinde K, et al. 2011. Biomimetic silicification of fibrous chitin from diatoms. *Chem Mater.* 23:2973–2978.
207. Wysokowski M, et al. 2015. Poriferan chitin as a versatile template for extreme biomimetics. *Polymers.* 7:235–265.
208. Park KE, Jung SY, Lee SJ, Min B-M, Park WH. 2006. Biomimetic nanofibrous scaffolds: preparation and characterization of chitin/silk fibroin blend nanofibers. *Int J Biol Macromol.* 38:165–173.
209. Hong M-S, et al. 2018. Biomimetic chitin–silk hybrids: an optically transparent structural platform for wearable devices and advanced electronics. *Adv Funct Mater.* 28:1705480.
210. Salameh C, et al. 2020. Origin of transparency in scattering biomimetic collagen materials. *Proc Natl Acad Sci.* 117:11947–11953.
211. Zheng W, Zhang W, Jiang X. 2010. Biomimetic collagen nanofibrous materials for bone tissue engineering. *Adv Eng Mater.* 12:B451–B466.
212. Minardi S, et al. 2017. Biomimetic collagen/elastin meshes for ventral hernia repair in a rat model. *Acta Biomater.* 50: 165–177.
213. McColgan J, McGookin EW. 2016. Coordination of multiple biomimetic autonomous underwater vehicles using strategies based on the schooling behaviour of fish. *Robotics.* 5:2.
214. Whittlesey RW, Liska S, Dabiri JO. 2010. Fish schooling as a basis for vertical axis wind turbine farm design. *Bioinspir Biomim.* 5:035005.
215. Pawlyn M. 2019. Biomimicry in architecture. London, UK: RIBA Publishing.
216. Volta A. 1800. XVII. On the electricity excited by the mere contact of conducting substances of different kinds. In a letter from Mr. Alexander Volta, F. R. S. Professor of Natural Philosophy in the University of Pavia, to the Rt. Hon. Sir Joseph Banks, Bart. K.B. P. R. S. Philos. Trans R Soc Lond. 90:403–431.
217. US20110200563 Calcium-mediated effects of coral and methods of use thereof. [accessed 2022 Sep 29]. https://patentscope.wipo.int/search/en/detail.jsf?docId=US73326595&_cid=P21-KYVKTA-62870-1.
218. Altschuler N. 2020. Optimized cage systems promoting bone repair and fusion. [accessed 2022 Sep 29]. <https://patents.google.com/patent/IL261820A/en>.
219. Altschuler N. 2013. Multi-phasic solid implants for tissue repair. [accessed 2022 Sep 29]. <https://patents.google.com/patent/WO2013150537A1/en>.
220. Altschuler N. 2014. Solid substrates for mitigating or preventing cell and tissue adhesion and vascularization. [accessed 2022 Sep 29]. <https://patents.google.com/patent/WO2014125477A1/pt-PT>.
221. Schlüter L, Mortensen L, Kørnøv L. 2020. Industrial symbiosis emergence and network development through reproduction. *J Cleaner Prod.* 252:119631.
222. Morris RL, Konlechner TM, Ghisalberti M, Swearer SE. 2018. From grey to green: efficacy of eco-engineering solutions for nature-based coastal defence. *Global Change Biol.* 24:1827–1842.
223. Jongman B, Gutierrez Goizueta G, Van Zanten B, Gonzalez Reguero B. 2021. Blue barriers: a nature-based solution to build resilience. [accessed 2022 Sep 29]. <https://blogs.worldbank.org/climatechange/blue-barriers-nature-based-solution-build-resilience>.
224. MarinLit. 2022. MarinLit, a database of marine natural products literature. [accessed 2022 Sep 29]. <https://marinlit.rsc.org>.
225. Carroll AR, Copp BR, Davis RA, Keyzers RA, Prinsep MR. 2021. Marine natural products. *Nat Prod Rep.* 38:362–413.
226. Haque N, Parveen S, Tang T, Wei J, Huang Z. 2022. Marine natural products in clinical use. *Mar Drugs.* 20:528.
227. Mayer AMS, Rodríguez AD, Taglialatela-Scafati O, Fusetani N. 2017. Marine pharmacology in 2012–2013: marine compounds with antibacterial, antidiabetic, antifungal, anti-inflammatory, antiprotazoal, antituberculosis, and antiviral activities; affecting the immune and nervous systems, and other miscellaneous mechanisms of action. *Mar Drugs.* 15: 273.
228. Molinski TF, Dalisay DS, Lievens SL, Saludes JP. 2009. Drug development from marine natural products. *Nat Rev Drug Discovery.* 8:69–85.
229. Papon N, Copp BR, Courdavault V. 2022. Marine drugs: biology, pipelines, current and future prospects for production. *Biotechnol Adv.* 54:107871.
230. Sigwart JD, Blasiak R, Jaspars M, Jouffray J-B, Tasdemir D. 2021. Unlocking the potential of marine biodiscovery. *Nat Prod Rep.* 38:1235–1242.
231. Gerwick WH, Moore BS. 2012. Lessons from the past and charting the future of marine natural products drug discovery and chemical biology. *Chem Biol.* 19:85–98.
232. Chalfie M, Tu Y, Euskirchen G, Ward WW, Prasher DC. 1994. Green fluorescent protein as a marker for gene expression. *Science.* 263:802–805.
233. Remington SJ. 2011. Green fluorescent protein: a perspective. *Protein Sci.* 20:1509–1519.
234. Ingham M, Schwartz GK. 2017. Cell-cycle therapeutics come of age. *J Clin Oncol.* 35:2949–2959.
235. Institut Océanographique. 2022. Models for science. Institut océanographique. [accessed 2022 Sep 29]. <https://www.oceano.org/en/resources/4-models-for-science/>.
236. Unterlass MM. 2017. Geomimetics and extreme biomimetics inspired by hydrothermal systems—what can we learn from nature for materials synthesis? *Biomimetics.* 2:8.
237. Baumgartner B, Bojdy MJ, Unterlass MM. 2014. Geomimetics for green polymer synthesis: highly ordered polyimides via hydrothermal techniques. *Polym Chem.* 5:3771–3776.
238. Kim T, et al. 2022. Geomimetic hydrothermal synthesis of polyimide-based covalent organic frameworks. *Angew Chem.* 134:e202113780.
239. Tyler PA. 2003. Ecosystems of the deep oceans. Amsterdam, Netherlands: Elsevier.
240. Rainey FA, Oren A. 2006. Extremophile microorganisms and the methods to handle them. In *Methods in microbiology*. vol. 35. Amsterdam, Netherlands: Elsevier. pp. 1–25.
241. Rothschild LJ, Mancinelli RL. 2001. Life in extreme environments. *Nature.* 409:1092–1101.
242. Glover A, Higgs N, Horton T. 2022. World register of deep-sea species (WoRDS). [accessed 2022 Sep 29]. <https://www.marine-species.org/deepsea/statistics.php>.

243. Levin L. 2021. IPCC and the deep sea: a case for deeper knowledge. *Front Clim.* 3: 720755.
244. Land MF. 2009. Biological optics: deep reflections. *Curr Biol.* 19:R78–R80.
245. Wagner H-J, Partridge JC, Douglas RH. 2019. Observations on the retina and 'optical fold' of a mesopelagic sabretooth fish, *Evermanella balbo*. *Cell Tissue Res.* 378:411–425.
246. Wagner H-J, Douglas RH, Frank TM, Roberts NW, Partridge JC. 2009. A novel vertebrate eye using both refractive and reflective optics. *Curr Biol.* 19:108–114.
247. Partridge JC, et al. 2014. Reflecting optics in the diverticular eye of a deep-sea barreleye fish (*Rhynchohyalus natalensis*). *Proc R Soc B Biol Sci.* 281:20133223.
248. Tamagawa T, et al. 2020. Multiplexing lobster-eye optics: a concept for wide-field X-ray monitoring. *J Astron Telesc Instrum Syst.* 6:025003.
249. Tichý V, et al. 2011. Tests of lobster eye optics for small space X-ray telescope. *Nucl Instrum Methods Phys Res Sect Accel Spectrometers Detect Assoc Equip.* 633:S169–S171.
250. Hudec R, Remisova K. 2017. Application of biomimetics in X-ray optics. In *EUV and X-ray optics: synergy between laboratory and space V.* vol. 10235. SPIE. pp. 23–31.
251. Remisova K, Hudec R. 2017. Application of biomimetics principles in space optics. In *International Conference on Space Optics—ICSO 2016.* vol. 10562. pp. 231–238. SPIE.
252. Trung HTD, Lee D, Nguyen TL, Lee H. 2021. Image formation by a biological curved mirror array of the fisheye in the deep-sea environment. *Appl Opt.* 60:5227–5235.
253. Haque O. 2019. Biomimicry-inspired design for daylighting through roof of multipurpose hall. *Bangladesh University of Engineering and Technology.* 162.
254. Wainwright O. 2014. Will the buildings of the future be grown underwater?. *The Guardian.* [accessed 2022 Sep 29]. <https://www.theguardian.com/artanddesign/architecture-design-blog/2014/feb/17/buildings-future-underwater-nature-architecture-michael-pawlyn-exhibition>.
255. Subramanian GR. 2010. Bionic passenger car. [accessed 2022 Sep 29]. <https://patentscope.wipo.int/search/en/detail.jsf?docId=IN211512310>.
256. Velasco-Hogan A, et al. 2019. On the nature of the transparent teeth of the deep-sea dragonfish, *Aristostomias scintillans*. *Mater.* 1:235–249.
257. UC San Diego. 2019. Researchers discover what makes deep-sea dragonfish teeth transparent. *ScienceDaily.* [accessed 2022 Sep 29]. <https://www.sciencedaily.com/releases/2019/06/190605133501.htm>.
258. Gerringer ME, Linley TD, Jamieson AJ, Goetze E, Drazen JC. 2017. *Pseudoliparis swirei* sp. nov.: a newly-discovered hadal snailfish (Scorpaeniformes: Liparidae) from the Mariana Trench. *Zootaxa.* 4358:161–177.
259. Li G, et al. 2021. Self-powered soft robot in the Mariana Trench. *Nature.* 591:66–71.
260. Fudge DS, Schorno S, Ferraro S. 2015. Physiology, biomechanics, and biomimetics of hagfish slime. *Annu Rev Biochem.* 84:947–967.
261. Yong E. 2019. No one is prepared for hagfish slime. *The Atlantic.* [accessed 2022 Sep 29]. <https://www.theatlantic.com/science/archive/2019/01/hagfish-slime/581002/>.
262. Baggaley K. 2017. Your guide to the practical uses of hagfish slime, glowworm glue, and other animal goo. *Popular Science.* [accessed 2022 Sep 29]. <https://www.popsci.com/animal-goo/>.
263. Linnaeus C. 1758. *Systema naturae*. Amsterdam: Nieuwkoop B. De Graaf.
264. WIPO. 2022. WIPO—Search International and National Patent Collections. [accessed 2022 Sep 29]. <https://patentscope.wipo.int/search/en/search.jsf>.
265. Böni LJ. 2018. *Biophysics and biomimetics of hagfish slime*. Zurich, Switzerland: ETH Zurich.
266. Dance A. 2016. Inner workings: will hagfish yield the fibers of the future? *Proc Natl Acad Sci.* 113:7005–7006.
267. Martins E, Rocha MS, Silva TH, Reis RL. 2019. Remarkable body architecture of marine sponges as biomimetic structure for application in tissue engineering. In Choi AH, Ben-Nissan B, editors. *Marine-derived biomaterials for tissue engineering applications*. Singapore: Springer. pp. 27–50.
268. Martins E, et al. 2021. Macro and microstructural characteristics of North Atlantic deep-sea sponges as bioinspired models for tissue engineering scaffolding. *Frontiers in Marine Science.* 7, p.613647.
269. Clarke SA, Walsh P, Maggs CA, Buchanan F. 2011. Designs from the deep: marine organisms for bone tissue engineering. *Biotechnol Adv.* 29:610–617.
270. Green DW, Ben-Nissan B, Yoon KS, Milthorpe B, Jung H-S. 2017. Natural and synthetic coral biomineralization for human bone revitalization. *Trends Biotechnol.* 35:43–54.
271. Wang X, Schröder HC, Feng Q, Draenert F, Müller WEG. 2013. The deep-sea natural products, biogenic polyphosphate (bio-polyp) and biogenic silica (bio-silica), as biomimetic scaffolds for bone tissue engineering: fabrication of a morphogenetically-active polymer. *Mar Drugs.* 11:718–746.
272. Falcucci G, et al. 2021. Extreme flow simulations reveal skeletal adaptations of deep-sea sponges. *Nature.* 595:537–541.
273. Fernandes MC, Aizenberg J, Weaver JC, Bertoldi K. 2021. Mechanically robust lattices inspired by deep-sea glass sponges. *Nat Mater.* 20:237–241.
274. Tavangarian F, Sadeghzade S, Davami K. 2021. A novel biomimetic design inspired by nested cylindrical structures of spicules. *J Alloys Compd.* 864:158197.
275. Müller WEG et al. 2008. Bioorganic/inorganic hybrid composition of sponge spicules: matrix of the giant spicules and of the comitalia of the deep sea hexactinellid *Monorhaphis*. *J Struct Biol.* 161:188–203.
276. Miserez A et al. 2008. Effects of laminate architecture on fracture resistance of sponge biosilica: lessons from nature. *Adv Funct Mater.* 18:1241–1248.
277. Salim NV, Jin X, Mateti S, Subhani K. 2021. Biomimetic hierarchical porous carbon fibers via block copolymer self-assembly. *Microporous Mesoporous Mater.* 321:111136.
278. Pinter G, Arbeiter F, Wiener J, Frank A, Kolednik O. 2018. Biomimetic design concepts for the pipe architecture of tomorrow. In *Plastic Pipes XIX Conference*; Las Vegas, NV.
279. Bohlin L, Sjögren M, Jonsson P, Göransson U. Antifouling agent. 2003. [accessed 2022 Sep 29]. <https://patents.google.com/patent/WO2003081199A2/en>.
280. Heinemann S, Worch H, Hanke T. 2009. US20110237552 Composite material consisting of a collagen matrix mineralised with silicate and calcium phosphate phases, method for the production and use thereof. [accessed 2022 Sep 29]. https://patentscope.wipo.int/search/en/detail.jsf?docId=US73363584&_cid=P21-KYVLBS-69193-1.
281. Denton EJ. 1971. Examples of the use of active transport of salts and water to give buoyancy in the sea. *Philos Trans R Soc Lond B Biol Sci.* 262:277–287.
282. Stewart KA. 2019. Understanding the effects of biotic and abiotic factors on sources of aquatic environmental DNA. *Biodivers Conserv.* 28:983–1001.

283. Murray L. 2012. Cameron sinks to the challenge. *Eng Technol*. 7:88–90.
284. Sigwart JD et al. 2019. Red Listing can protect deep-sea biodiversity. *Nat Ecol Evol*. 3:1134–1134.
285. Kapsali V. 2013. Biomimetic approaches to the design of smart textiles for protection. In Chapman RA, editor. *Smart textiles for protection*. Sawston, UK: Woodhead Publishing, pp. 214–226.
286. Yao H et al. 2010. Protection mechanisms of the iron-plated armor of a deep-sea hydrothermal vent gastropod. *Proc Natl Acad Sci*. 107:987–992.
287. Islam MR, Jeong YT, Ryu YJ, Song CH, Lee YS. 2009. Isolation, identification and optimal culture conditions of *Streptomyces albidoflavus* C247 producing antifungal agents against *Rhizoctonia solani* AG2-2. *Mycobiology*. 37:114–120.
288. Yan L-L et al. 2010. Antimycin A18 produced by an endophytic *Streptomyces albidoflavus* isolated from a mangrove plant. *J Antibiot (Tokyo)*. 63:259–261.
289. Swiontek Brzezinska M, Jankiewicz U, Burkowska A. 2013. Purification and characterization of *Streptomyces albidoflavus* antifungal components. *Appl Biochem Microbiol*. 49:451–457.
290. Xu Y et al. 2010. Potent antifouling compounds produced by marine *Streptomyces*. *Bioresour Technol*. 101:1331–1336.
291. Davidson BS. 1995. New dimensions in natural products research: cultured marine microorganisms. *Curr Opin Biotechnol*. 6:284–291.
292. Qian P, et al. 2010. Antifouling furan-2-one derivatives. [accessed 2022 Sep 29]. <https://patents.google.com/patent/CN102105542B/en>.
293. Reed JK. 2002. Comparison of deep-water coral reefs and lithohierms off southeastern USA. *Hydrobiologia*. 471:57–69.
294. Fosså JH, Mortensen PB, Furevik DM. 2002. The deep-water coral *Lophelia pertusa* in Norwegian waters: distribution and fishery impacts. *Hydrobiologia*. 471:1–12.
295. Järnegren J, Brooke S, Jensen H. 2017. Effects of drill cuttings on larvae of the cold-water coral *Lophelia pertusa*. *Deep Sea Res Part II Top Stud Oceanogr*. 137:454–462.
296. Baussant T, Nilsen M, Ravagnan E, Westerlund S, Ramanand S. 2018. Effects of suspended drill cuttings on the coral *Lophelia pertusa* using pulsed and continuous exposure scenarios. *J Toxicol Environ Health A*. 81:361–382.
297. AU2015308950 Biosensor device, system and method for monitoring a deep-water sea-floor. [accessed 2022 Sep 29]. https://patentscope.wipo.int/search/en/detail.jsf?docId=AU194375220&_cid=P21-KYVM2L-78655-1.
298. Ehrlich H, et al. 2006. Biomaterial structure in deep-sea bamboo coral (Anthozoa: Gorgonacea: Isididae): perspectives for the development of bone implants and templates for tissue engineering. *Mat-wiss u Werkstofftech*. 37:552–557.
299. The common sea mussel will protect us against rust. KTH. [accessed 2022 Sep 29]. <https://www.kth.se/en/om/nyheter/centrala-nyheter/blamusslor-ska-skydda-mot-rost-1.66320>.
300. Kim YS, Smoak MM, Melchiorri AJ, Mikos AG. 2019. An overview of the tissue engineering market in the United States from 2011 to 2018. *Tissue Eng Part A*. 25:1–8.
301. Underwater Robotics Market Size Worth \$6.74 Billion By 2025. [accessed 2022 Sep 29]. <https://www.grandviewresearch.com/press-release/global-underwater-robotics-market>.
302. Howell KL, et al. 2020. A blueprint for an inclusive, global deep-sea ocean decade field program. *Front Mar Sci*. 7.
303. Claudet J, et al. 2020. A roadmap for using the UN decade of ocean science for sustainable development in support of science, policy, and action. *One Earth*. 2:34–42.
304. Arico S, et al. 2020. Global ocean science report 2020—charting capacity for ocean sustainability. Paris, France: UNESCO.
305. IPBES, 2022. Summary for policymakers of the methodological assessment of the diverse values and valuation of nature of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services (IPBES). [accessed 2022 Sep 29]. <https://ipbes.net/the-values-assessment>.
306. United Nations, 2021. The Second World Ocean Assessment. New York, USA: United Nations.
307. Leggat WP, et al. 2019. Rapid coral decay is associated with marine heatwave mortality events on reefs. *Curr Biol*. 29:2723–2730.e4.
308. Johnstone JW, Waller RG, Stone RP. 2021. Shallow-emerged coral may warn of deep-sea coral response to thermal stress. *Sci Rep*. 11:22439.
309. Roessger J, Claudet J, Horta e Costa B. 2022. Turning the tide on protection illusions: the underprotected MPAs of the “OSPAR Regional Sea Convention”. *Mar Policy*. 142:105109.
310. Norström AV, et al. 2016. Guiding coral reef futures in the Anthropocene. *Front Ecol Environ*. 14:490–498.
311. PROBLUE: The World Bank’s Blue Economy Program. 2022. World Bank. [accessed 2022 Sep 29]. <https://www.worldbank.org/en/programs/problue>.
312. Sustainable Blue Economy | Commonwealth. 2022. [accessed 2022 Sep 29]. <https://thecommonwealth.org/bluecharter/sustainable-blue-economy>.
313. Silver JJ, Gray NJ, Campbell LM, Fairbanks LW, Gruby RL. 2015. Blue economy and competing discourses in international oceans governance. *J Environ Dev*. 24:135–160.
314. Anon. 2014. Blue economy concept paper. [accessed 2022 Sep 29]. https://wedocs.unep.org/bitstream/handle/20.500.11822/11129/unep_swio_sm1_inf11_blue_economy.pdf.
315. Aisi R 2011. Statement by H.E. Mr. Robert G Aisi, permanent representative of Papua New Guinea to the UN on behalf of the Pacific Small Island Developing States. [accessed 2022 Sep 29]. <https://sustainabledevelopment.un.org/content/documents/18805PSIDS.pdf>.
316. UN Environment Programme Finance Initiative. 2021. Turning the tide: how to finance a sustainable ocean recovery—A practical guide for financial institutions. Geneva: UN Environment Programme.